Cold Atmospheric Plasma and Plasma-Activated Medium: Antitumor Cell Effects with Inherent Synergistic Potential

Georg Bauer

Institute of Virology, Medical Center and Faculty of Medicine, University of Freiburg, Germany; Tel.: 0049 761 203 6618, Fax: 0049 761 203 6562, E-mail: georg.bauer@uniklinik-freiburg.de

ABSTRACT: Nitrite and H₂O₂, long-lived molecular species from cold atmospheric plasma (CAP) and plasma-activated medium (PAM), reach tumor target cells in vitro and in vivo. Through several steps, the interaction between nitrite and H₂O₂ leads to generation of singlet oxygen (¹O₂). ¹O₂, then interacts with a specific biochemical switchboard on tumor cells that is composed of catalase, superoxide dismutase (SOD), first aptosis signal (FAS) receptor, and nicotinamide adenine dinucleotide phosphate (NADPH) oxidase. As a result, local inactivation of catalase by minute concentrations of primary singlet oxygen opens a strong autoamplificatory sustained process of secondary singlet oxygen generation and catalase inactivation. This process is driven by tumor cell-specific NADPH oxidase-1 and spreads within the tumor cell population. The concerted action of singlet oxygen interaction with catalase, SOD, and FAS receptor causes an efficient mode of synergistic interaction. Defined reactive oxygen and reactive nitrogen species (ROS/RNS) such as H₂O₂ and nitrite have multiple functions in this process. Catalase-mediated oxidation of nitrite enhances generation of nitrogen dioxide, which is rate limiting for singlet oxygen generation. Before singlet oxygen-mediated inactivation of catalase and, subsequently, reactivated intercellular ROS/RNS signaling can activate the mitochondrial pathway of apoptosis, counteraction of glutathione to lipid peroxidation must be abrogated through aquaporin-mediated influx of H₂O₂ into cells. CAP- and PAM-dependent immunogenic cell death triggers a strong immune response that finalizes antitumor action in vivo. Thus, the high efficiency of CAP and PAM seem to depend on concerted action of several dominant steps and their autoamplificatory potential.

KEY WORDS: cold atmospheric plasma, plasma-activated medium, singlet oxygen, catalase, immunogenic cell death

I. INTRODUCTION

The complexity of the physics and chemistry of cold atmospheric plasma (CAP) and plasma-activated medium (PAM) is paralleled by their striking biological effects, ranging from antimicrobial action to stimulating wound healing and impressive antitumor effects *in vitro* and *in vivo*. ^{1–19} CAP and PAM represent a reactive oxygen and reactive nitrogen species (ROS/RNS)-dependent system that may be applicable to an impressively wide range of tumors that are derived from different tissues and show a high degree of selective action in most systems studied. CAP and PAM are unique with respect to their mode of initial interaction with tumor cells and tumor tissue: Initial interaction of ROS and RNS with tumor cells occurs primarily on the outside of the cells. No other experimental antitumor system has this specific feature. Even photodynamic therapy that uses the power of one defined initial ROS, that is, singlet oxygen, does not apply its key molecule to the

outside of malignant cells, because the photosensitizer is given sufficient time to enter cells before illumination triggers intracellular production of singlet oxygen.

Novel insights into the chemical biology of malignant cells during tumor progression, rich literature on CAP- and PAM-dependent chemistry and biology, and data on the interaction of defined ROS and RNS with malignant and nonmalignant cells allow investigators to propose a potential mechanism of CAP- and PAM-dependent antitumor effects. ^{20,21} The high efficiency and impressive selectivity of CAP- and PAM-mediated antitumor effects have served to inspire us to search for evidence-based potential synergistic effects that may be inherent to CAP and PAM action. The biochemical basis of these synergistic effects is discussed in this conceptional review. Detailed knowledge about these synergistic effects may be valuable for further optimization of CAP or PAM treatment. This may become especially relevant under conditions allowing only the application of lower doses, such as future endoscopic application of CAP or PAM to tumors in internal organs.

II. CONCEPTIONAL REVIEW

A. Basic Principles of CAP and PAM Action

1. Physical Plasma and Plasma Medicine

The gaseous and liquid phases of CAP are a rich source of radical and nonradical ROS/RNS.^{1–19} Due to variable lifetimes, ranges of action, and multiple potentials of interaction, these species represent a unique scenario in ROS/RNS chemical biology. The treatment of media with CAP results in generation of PAM that maintains the major biological effects of CAP, although it only contains long-lived species from CAP such as nitrite, nitrate, and H₂O₂.^{22–25}

The study of the effects of exogenous ROS/RNS in CAP and PAM on biological systems has opened the exiting new field of plasma medicine. ^{2,3,6,8,9,26–35} CAP and PAM cause impressive antibacterial ^{36–38} and antiviral effects ³⁹ and benefit wound healing ^{40–43} and treatment of actinic keratosis. ^{44–46} A particular focus is on promising *in vitro* and *in vivo* antitumor effects on a broad variety of tumor systems. ^{8,9,12,29,30,47–55} Clinical application of CAP in tumor therapy yielded the first encouraging results without severe side effects. ^{31–34,56} Although CAP and PAM seem to target a rather general principle regarding tumor cells, the mechanisms underlying their antitumor action are still a matter of scientific debate.

2. CAP- and PAM-Mediated Antitumor Effects

In most studies, antitumor effects of CAP and PAM are found to be selective with respect to the malignant phenotype of target cells *in vitro* and *in vivo*.^{22,57–68} Only a few reports claim to have found nonselective apoptosis-inducing effects of CAP or PAM.^{69–73} Further studies, including those for standardizing CAP and PAM doses and composition,

may clarify whether this discrepancy might simply be explained by a conceivable correlation between doses and selectivity or nonselectivity of action.

a. Initial Effects of CAP and PAM on Tumor Cells

General agreement exists that initial CAP effects on tumor cells are mediated by CAP-derived ROS/RNS and amplified by intracellular ROS. 8,9,13,29,30,47,48,50,51,52–55,74 Because apoptosis is induced in areas of treated tumors that are not directly reached by constituents of CAP, a self-perpetuating signaling system was postulated. 12,13,20,47,53 Tumor treatment with CAP and PAM therefore seems to have a strong conceptual and mechanistic overlap with other ROS/RNS-dependent treatments, such as photodynamic therapy and modulating the endogenous NO level of tumor cells for subsequent ROS/RNS-driven processes that lead to selective apoptosis induction in tumor cells. 75,76 Therefore, the well-characterized ROS/RNS composition of CAP and PAM and the chance to modulate their composition offer great analytical potential for research on antitumor action of CAP and PAM and their application. Analogous to photodynamic therapy that requires interaction between an initial singlet oxygen–dependent step and a subsequent immunological process, 77 CAP and PAM action may also be connected to strong immunological processes.

b. Immunogenic Cell Death Induced by CAP and PAM

It seems that CAP- and PAM-dependent selective apoptosis induction in tumor cells has characteristics of immunogenic cell death; that is, stimulation of dendritic cells that thus provokes subsequent activation of cytotoxic T cells. ^{78–85} In addition, CAP gives rise to activation of cytotoxic macrophages. ⁸⁶ The findings that CAP- and PAM-induced cell death may trigger an additional strong and selective immunological antitumor effect, that is far ranging within the tumor and the body, is in perfect agreement with the general concept that immunological cell death is a necessary element in finalizing tumor treatment by various antitumor agents. ^{77,87–91}

3. Common Principles of CAP and PAM Action in Tumor Cells

It is intriguing that PAM mimics antimicrobial and antitumor effects of CAP, $^{22-24,38,58,92-98}$ although its composition seems to be restricted to H_2O_2 , nitrite, and nitrate. $^{22-25}$ However, it is important to consider that the spectrum of species contained in the liquid phase of CAP that can actually reach tumor cells *in vitro* or *in vivo* depends essentially on volume and composition of medium or biological material above or around the target cells. Highly reactive CAP species may likely be filtered out before they can react with target cells, whereas long-lived and far-ranging species such as H_2O_2 , nitrite, and nitrate have a good chance of reaching target cells. Model experiments show that CAP-derived molecular species that can actually reach a tumor *in vivo* may be the same as those found

in PAM. $^{99-103}$ How can relatively simple species such as H_2O_2 , nitrite, and nitrate mediate such efficient, complex, and selective effects on tumor cells?

4. Divergent Working Hypotheses for CAP and PAM Action

a. Classical Concept for CAP and PAM: Direct Action on Target Cells

Working hypotheses of most investigators were previously based on assumptions that the molecular species acting selectively on tumor cells are (1) already present in CAP or PAM itself, and (2) present at sufficiently high concentrations to allow direct induction of apoptotic or necrotic cell death. However, this conceptional approach has not yet led to the establishment of an experimentally determined concise model for CAP- and PAM-dependent signaling chemistry that is in perfect agreement with existing central biological and chemical observations.

 $\rm H_2O_2$ in particular had often been assumed to act as a direct, selective apoptosis inducer of CAP and PAM. However, this assumption disagrees with experimental assessments showing that tumor cells are less sensitive to exogenous $\rm H_2O_2$ than nonmalignant cells. 104,105 This is due to the expression of membrane-associated catalase on the tumor cell membrane. $^{20,21,105-114}$ Therefore, $\rm H_2O_2$ by itself cannot cause selective apoptosis induction in tumor cells.

b. Novel Concept for CAP and PAM Action: Triggering Specific Cellular and Intercellular Signaling Pathways

An alternative model to explain selective antitumor action of CAP and PAM was proposed by Bauer and Graves. This model is based on the concept that an initially low concentration of singlet oxygen contained in or generated by CAP and PAM is insufficient to induce cell death directly but triggers tumor cells to generate higher concentrations of secondary singlet oxygen in an autoamplificatory process. This leads to inactivation of protective catalase in the membrane of the initially targeted cell, followed by a bystander signaling-like spread of catalase inactivation on neighboring cells. Finally, this allows reactivation of intercellular ROS/RNS-dependent apoptosis-inducing signaling within the population of tumor cells. This model is based on experimental evidence that was obtained using multiple approaches in which singlet oxygen—dependent effects on tumor cells were studied. 75,76,109,115-117

The initially determining switching-on effect by CAP- or PAM-derived singlet oxygen allows strong subsequent and continuous generation of secondary singlet oxygen by tumor cells. This is based on their active nicotinamide adenine dinucleotide phosphate (NADPH) oxidase (NOX) and NO synthase (NOS), with H_2O_2 and peroxynitrite acting as intermediates. ^{109,115–117} Selectivity of CAP and PAM action for tumor cells is thus warranted by the (1) initial singlet oxygen/catalase interaction, (2) potential of tumor cells (but not nonmalignant cells) to drive NOX1- and NOS-dependent generation of secondary singlet oxygen, and (3) NOX1-driven intercellular ROS/RNS-dependent apoptosis-inducing signaling after catalase inactivation.

c. Role of Aquaporins during CAP and PAM Action

Yan et al. presented strong evidence for a dominant controlling function of aquaporins related to the action of CAP and PAM. These investigators also suggested that aquaporins are primary interaction partners for CAP- or PAM-derived H₂O₂. In their model, selectivity of CAP and PAM action for tumor cells is explained by the higher concentration of aquaporins in tumor cell membranes. However, this model did not take into account that membrane-associated catalase on tumor cells may decompose exogenous H₂O₂ before it can pass through aquaporins. Therefore, Bauer suggested that the contribution of aquaporins to the sensitization of tumor cells by CAP and PAM requires preceding catalase inactivation. Recently, this concept was experimentally verified in kinetics studies showing that aquaporin action was required *after* catalase inactivation due to singlet oxygen derived from CAP or generated by PAM (Bauer, manuscript in preparation). Because glutathione depletion substitutes for aquaporin function, the significance of aquaporin-dependent influx of H₂O₂ seems to be related to sensitization of tumor cells for exogenous ROS effects through reduced endogenous glutathione levels.

Completed experimental work using a bottom-up experimental approach with compounds that are found in PAM confirmed central predictions of our previous models. A portion of these data was presented at the 7th International Conference on Plasma Medicine (ICPM-7) in Phildadelphia.

B. Chemical Biology of CAP and PAM Interaction with Tumor Cell Surface

Evaluation of possible interactions of CAP and PAM with the surface of tumor cells requires (1) detailed knowledge of the complex nature of the membrane of tumor cells, ^{21,76,108,111,112} (2) composition of CAP and PAM, ^{1–19} (3) free diffusion path length of CAP- and PAM-derived ROS/RNS, and (4) scavenging effect of biological materials for CAP- and PAM-derived molecular species. ^{99–103}

1. Biochemistry of Malignant and Nonmalignant Cell Surfaces

Essential aspects of the surface of tumor cells compared to nonmalignant cells are shown in Fig. 1. These central aspects include active NOX1 (the hallmark of malignant cells)^{120–124,111,112} and membrane-associated catalase that protects tumor cells from NOX1-driven intercellular ROS/RNS signaling. Because NOX-derived superoxide anions have the potential to inhibit catalase,^{125–128} catalase function requires the parallel expression of membrane-associated superoxide dismutase (SOD) that reduces the concentration of superoxide anions. Whereas the concentration of membrane-associated catalase is sufficiently high to completely prevent intercellular ROS/RNS signaling,^{105,108} SOD concentration on the surface does not allow complete removal of free superoxide anions but ensures catalase function by reducing concentration of superoxide anions to a level that does not lead to inhibition of catalase.^{110,129}

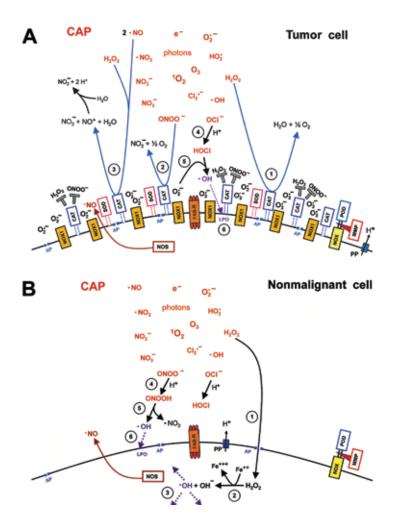


FIG. 1: Interaction of CAP with tumor cells and nonmalignant cells. (A) Tumor cells. The membrane of bona fide tumor cells contains superoxide anion–generating NOX1 and dual oxidase (DUOX), which consists of a peroxidase domain (POD) and a NOX domain. POD is removed from DUOX through matrix metalloprotease (MMP) action. Catalase (CAT) is attached to the outside of the membrane and decomposes the H₂O₂ and peroxynitrite that is generated by tumor cells. SOD that is attached to the membrane prevents superoxide anion–mediated inhibition of catalase. The membrane contains aquaporins (AP), proton pumps (PP), and FAS receptor. CAP-derived H₂O₂ (1) and peroxynitrite (2) is decomposed by catalase, whereas NO is oxidized by catalase (3). Oxidation of NO requires the formation of compound I through catalase/H₂O₂ interaction. Protonation of hypochloride anions leads to the formation of HOC1 (4). The interaction of HOC1 with superoxide anions (5) causes generation of hydroxyl radicals that induce lipid peroxidation (6). (B) Nonmalignant cells. H₂O₂ passes the membrane through aquaporins (1) and is subject to intracellular Fenton chemistry (2),(3), resulting in damage and apoptosis-inducing hydroxyl radicals. Peroxynitrite causes lipid peroxidation (LPO) and apoptosis through (4)–(6).

Further membrane elements of high significance for signaling effects are aquaporins that control the influx of H₂O₂ into cells and proton pumps that allow conversion of peroxynitrite to peroxynitrous acid in close vicinity to the membrane.

2. Conceivable Interactions between CAP and PAM ROS/RNS and the Cell Surface

CAP contains a plethora of defined ROS/RNS, electrons, and photons, as summarized in Fig. 1(A).¹⁻¹⁹ Biological material surrounding the tumor and the layer of medium above tumor cells *in vitro* most likely consumes most of the highly reactive species derived from CAP or generated through CAP/liquid-phase interaction. This process thus selects for a few long-lived species. Therefore, CAP-derived electrons, photons, hydroxyl radicals, and superoxide anions seem to have no chance to reach tumors *in vivo* or tumor cells *in vitro* at concentrations that might potentially induce biological effects.⁹⁹⁻¹⁰³

Because the surface of tumor cells expresses catalase and SOD, CAP- or PAM-derived species with strong apoptosis-related potential such as H₂O₂, peroxynitrite, or NO are efficiently decomposed (H₂O₂, peroxynitrite) or oxidized (NO) by tumor cell protective catalase. ^{21,108,124} Thus, they have no chance to induce apoptosis in tumor cells, unless they are applied at very high concentrations. Hypochloride anions are potentially able to generate HOCl after approaching proton pumps in the membrane, followed by hydroxyl radical generation after HOCl/superoxide anion interaction. ¹²⁴ But HOCl also likely reacts with biological material surrounding the tumor. In addition, its apoptosis-inducing reaction does not have obvious potential to trigger self-perpetuation of the process. Therefore, the role of HOCl is rather limited for triggering a minor apoptotic response. This limitation might also be due to the absence of HOCl catalase inhibition, preventing influx of H₂O₂ into tumor cells. Therefore, the counteraction of intracellular glutathione peroxidase-4/glutathione for lipid peroxidation impedes the effect of HOCl-derived hydroxyl radicals.

Because nonmalignant cells do not express NOX1, and in particular have no catalase or SOD on their surface [Fig. 1(B)], CAP-derived hydrogen peroxide and peroxynitrite at sufficiently high concentrations may induce apoptosis in nonmalignant cells. ^{104,105} Because nonmalignant cells are more highly affected by exogenous H₂O₂ and peroxynitrite than are catalase-bearing tumor cells, the concentration of these two species in CAP and PAM must be below an apoptosis-inducing concentration for nonmalignant cells, when CAP- and PAM-mediated selective action toward tumor cells and sparing of nonmalignant accompanying cells would occur. Recent experimental evidence (Bauer, manuscript in preparation) shows that H₂O₂ concentrations that are too low to induce apoptosis in nonmalignant cells are sufficient to allow enough singlet oxygen generation to trigger cascades that lead to tumor cell death when combined with nitrite. Under these conditions, generation of low concentrations of singlet oxygen is ensured and seems to drive the subsequent biochemical steps that lead to tumor cell apoptosis.

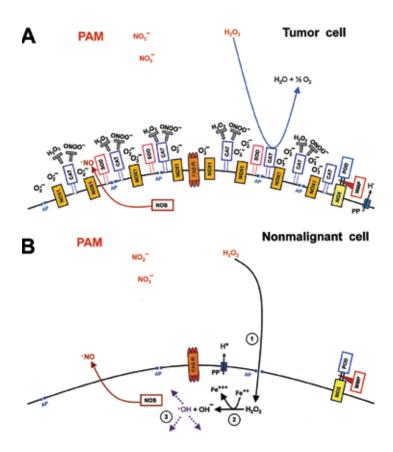


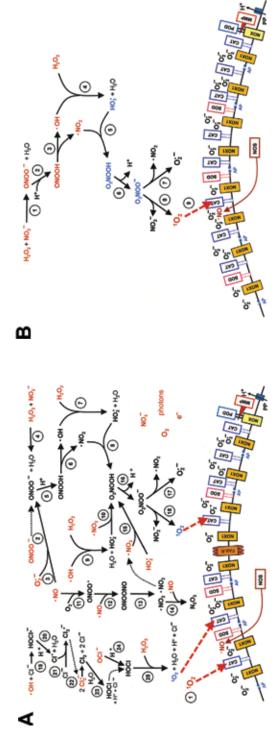
FIG. 2: Interaction of PAM with tumor cells and nonmalignant cells. PAM essentially contains H_2O_2 , nitrite, and nitrate. H_2O_2 is decomposed by membrane-associated catalase of tumor cells (A) and may induce apoptosis in nonmalignant cells through (1)–(3) (B), provided that it is present at sufficiently high concentrations. Selective apoptosis induction in tumor cells therefore requires the concentration of H_2O_2 to be too low to affect nonmalignant cells, and the reaction between H_2O_2 and nitrite must lead to generation of singlet oxygen, as outlined in Fig. 3.

It is obvious in Fig. 2 that H₂O₂, which is contained in PAM, is most likely decomposed by tumor cells, unless its concentration is extremely high. Very high concentrations of H₂O₂ can lead to SOD inactivation, followed by superoxide anion–mediated inhibition of neighboring catalase and onset of singlet oxygen–generation of H₂O₂.¹¹⁷ However, the high concentrations of H₂O₂ that are required for these effects do not allow selective apoptosis induction in tumor cells, because nonmalignant cells are affected by H₂O₂ even more strongly than tumor cells.¹⁰⁵ Therefore, we can conclude that establishment of a selective antitumor effect requires that H₂O₂ in PAM must be below an apoptosis-inducing concentration for nonmalignant cells. It is also obvious that the biological effect of PAM on tumor cells requires generation of a signaling species from its constituents to achieve specific apoptosis induction in tumor cells.

3. Signaling Effects through 10, Derived from CAP and PAM

Chemical biological effects of singlet oxygen that are generated by an illuminated photosensitizer shows that extracellular singlet oxygen efficiently mediates apoptosis induction in tumor cells, without affecting their nonmalignant counterparts.¹¹⁵ The selectivity of apoptosis induction by singlet oxygen is strictly dependent on its generation outside of the cells; it was lost when singlet oxygen was generated inside cells.115 It is thus safe to assume that as singlet oxygen that is derived from CAP or generated by PAM approaches target cells from the outside, it may exert the same specific antitumor effects as those established in model experiments with an extracellular photosensitizer. Figure 3(A) illustrates that singlet oxygen that is contained in CAP has the potential to cause local inactivation of a few catalase molecules on the surface of tumor cells, in accordance with recent findings. 115,130,131 Figure 3(A) also shows that the CAP liquid phase, in addition to singlet oxygen, contains various constituents that may lead to the generation of further singlet oxygen molecules through multiple, interconnected pathways. It becomes obvious that the sequence of reactions starting with reaction (4) in Fig. 3(A) may create a pathway that causes PAM to generate ¹O₂ through the scheme shown in Fig. 3(B). The potential relevance of reactions (1)–(3) in Fig. 3(B) had been already recognized by Girard et al., ²⁴ Kurake et al., ²³ and Jablonowski and von Woedtke. 132 Due to the relatively low reaction rate of (1), the importance of this scenario was initially underestimated by the plasma community. In light of the novel concept that PAM generates signaling molecules rather than directly damaging tumor cells, the slow reaction 1 can rather be seen as an advantage, because it ensures long-lasting generation of low concentrations of signaling molecules. It also ensures the availability of H₂O₂, for (4) after (1) is accomplished. Recent experimental work (Bauer, manuscript in preparation) has confirmed the reaction scheme for PAM action, as shown in Fig. 3(B).

Multiple completed model experiments that used either direct application of exogenous singlet oxygen¹¹⁵ or NO-mediated inhibition of catalase^{75,109,116} demonstrate that local inactivation of membrane-associated tumor cell catalase provokes a subsequent autoamplificatory generation of secondary singlet oxygen that is driven by free H₂O₂ and peroxynitrite at the site of inactivated catalase. Due to the activity of NOX1 and NOS, tumor cells continuously generate H₂O₂ and peroxynitrite. Thus, the sustained generation of secondary singlet oxygen is ensured. This is followed by inactivation of large amounts of tumor cell protective catalase molecules on the tumor cell membrane. As a result, intercellular apoptosis-inducing ROS/RNS signaling is subsequently reactivated. We may therefore conclude that low concentrations of singlet oxygen from CAP or PAM are sufficient to trigger secondary singlet oxygen generation and catalase inactivation in an autoamplificatory mode. This scenario is summarized in Fig. 4(A) and has been confirmed by direct experimental approach (Bauer, manuscript in preparation). In contrast to tumor cells, nonmalignant cells are unaffected by extracellular singlet oxygen at low to moderate concentrations, because they lack both catalase and NOX1 [Fig. 4(B)].



In addition, most of the reactive species contained in CAP (labeled in red) have the chemical potential to contribute to generation of singlet oxygen. Peroxynitrite that is present in CAP (2) or generated through the interaction between superoxide anions and NO (3) or between FIG. 3: Singlet oxygen in CAP and PAM. (A) Multiple sources of singlet oxygen in CAP. CAP contains singlet oxygen (10.) (reaction 1). H,O, and nitrite (4) may lead to the formation of peroxynitric acid (O,NOOH) through (6)–(8), involving CAP-contained H,O,. Peroxynitric acid may alternatively be generated through (9), (10), or (15). Reaction (15) requires NO, to be present in CAP or generated through oxidation of NO (11)+(14). Peroxynitric acid leads to the formation of singlet oxygen through (16) and (18). Reactions (19)+(25) summarize alternative pathways for the generation of HOCl and singlet oxygen generation through the interaction between H,O, and HOCl. Singlet oxygen from all of these pathways has the potential to inactivate catalase through reaction with histidine at the active center, provided that it is generated sufficiently close to the enzyme. (B) Singlet oxygen generation in PAM. H,O, and nitrite are sufficient to generate singlet oxygen through (1)–(9), in analogy to the reaction described for CAP (4)–(8),(16)–(18) in (A). In this way, PAM has the potential for local nactivation of catalase molecules by singlet oxygen.

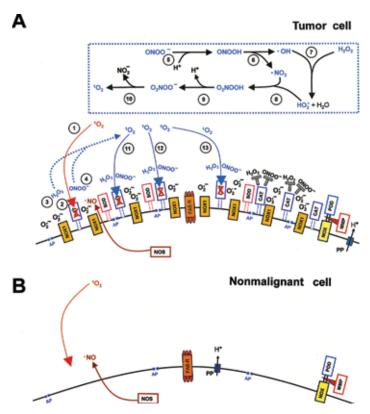


FIG. 4: Singlet oxygen from CAP or PAM triggers the generation of secondary singlet oxygen selectively by tumor cells. (A) Tumor cells. Singlet oxygen from CAP or PAM leads to local inactivation of catalase, followed by free H2O2 and peroxynitrite in close vicinity of the inactivated enzyme (1). Free H2O2 (3) and peroxynitrite (4) allow for the formation of secondary singlet oxygen (details in [5]–[10]). Secondary singlet oxygen causes inactivation of further catalase molecules (11)–(13) that will then allow further generation of singlet oxygen in an autoamplificatory process. It is important to note that singlet oxygen generation at the site of inactivated catalase is continuously propagated through a new supply with H2O2 and peroxynitrite that are derived from NOX1 and NOS. (B) Nonmalignant cells. Singlet oxygen from CAP or PAM finds no biologically relevant target structure on nonmalignant cells and therefore produces no effect (at up to moderately high concentrations).

4. Additional Potential Signaling Elements

Ozone has been found to inactivate catalase, ^{133,134} so CAP-derived ozone may potentially contribute to initial inactivation of tumor cell protective catalase. This would lead to generation of secondary singlet oxygen, analogous to the scenario triggered by initial singlet oxygen. Therefore, ozone might act in concert with singlet oxygen to induce the same signaling process, provided it can reach malignant target cells at sufficiently high concentrations.

Free electrons from CAP are not likely to reach the surface of tumors *in vivo* or tumor cells *in vitro* that are covered by a layer of medium. Therefore, electron-dependent reduction of catalase compound I (CATFe^{IV}= O·+) to compound II (CATFe^{IV}= O), followed by H_2O_2 —mediated generation of the terminally inactive compound III (CATFe^{III}O₂·-), is solely a theoretical possibility, although catalase inactivation through electron transfer has been observed in electron donors such as methyldopa. 116

C. Potential Synergistic Effects Inherent to CAP and PAM

1. Effects Induced by Singlet Oxygen

a. Synergistic Effects through Parallel Singlet Oxygen-Dependent Inactivation of Membrane-Associated Catalase and SOD

Due to its higher concentration on the cell membrane, catalase is more likely to react with the limited concentration of singlet oxygen derived from CAP or PAM than that from SOD. However, as soon as secondary singlet oxygen is generated through the complex interaction between H₂O₂ and peroxynitrite at the site of inactivated catalase, chances of singlet oxygen-dependent inactivation of SOD obviously increase [Fig. 5(A)]. 130,131 This then results in a local increase in concentration of NOX-1-derived superoxide anions at the site of inactivated SOD. Because superoxide anions can inhibit catalase, 125-128 superoxide anion-dependent inhibition of catalase in the neighborhood of inactivated SOD should then trigger higher generation of secondary singlet oxygen through the reaction between free H₂O₂ and peroxynitrite. One might assume that singlet oxygen–dependent inactivation of SOD thus merely contributes additively to the biochemical effects of inactivated catalase. But experimental analysis of specific neutralizing single-domain antibodies directed toward either catalase or SOD showed that parallel targeting of SOD and catalase causes a remarkable synergistic effect.¹¹⁰ Compared to single application of antibodies, the necessary concentration of the two antibodies used in the synergy experiments could be reduced to < 1% of the value required to achieve the same effect with single antibodies. It is justified to assume that parallel inactivation of catalase and SOD by singlet oxygen would cause a similar synergistic effect as that for parallel targeting with neutralizing antibodies. This mechanism may therefore significantly contribute to the synergistic potential that is inherent in CAP and PAM and at least partially explain the striking biological efficiency.

b. Synergistic Effects through Singlet Oxygen–Mediated Activation of First Apoptosis Signal (FAS) Receptor

Because FAS receptor can be activated by singlet oxygen (even in the absence of a genuine FAS ligand),¹³⁵ secondary singlet oxygen generated after initial CAP- or PAM-dependent singlet oxygen effects on tumor cell catalase can very likely have a chance to activate FAS receptor in target cells [Fig. 5(B)]. Usually, tumor cells do not express

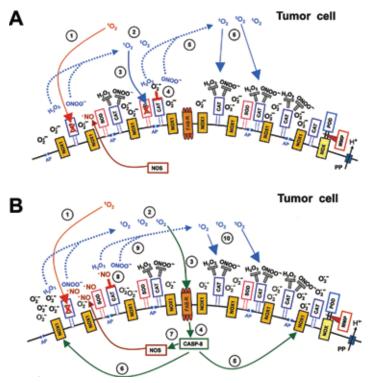


FIG. 5: Additional targets for secondary singlet oxygen on tumor cells. (A) Targeting of SOD. Secondary singlet oxygen (2) generated after the initial action of CAP- or PAM-derived primary singlet oxygen (1) may hit membrane-associated SOD and inactivate the enzyme through interaction with histidine at its active center. ^{130,131} As a consequence, the concentration of NOX-derived superoxide anions may increase in close vicinity to the inactivated enzyme and cause inhibition of neighboring catalase molecules. ^{125–128} This then triggers further amplification of secondary singlet oxygen generation. (B) Targeting of FAS receptor. Secondary singlet oxygen may activate FAS receptor (2),(3). ¹³⁵ As a consequence, caspase-8 is activated by the receptor and enhances activities of NOX1 and NOS (4)–(7). High local concentrations of NO may reversibly inhibit catalase (8). This allows further continuous generation of secondary singlet oxygen (9),(10).

sufficient FAS receptor to allow direct death receptor—dependent cell death through the canonical FAS pathway.⁷⁵ However, FAS receptor present even at low concentrations on tumor cells causes a dramatic increase in NOX1 and NOS activities.^{75,116,136–138} This leads to a valuable synergistic effect on singlet oxygen generation and subsequent catalase inactivation. Synergistic enhancement does not occur at very low concentrations of initially present singlet oxygen.¹¹⁵ Likewise, intercellular apoptosis-inducing ROS/RNS signaling after inactivation of catalase may take advantage of the increased concentrations of superoxide anions, NO, and their derivatives after singlet oxygen—mediated activation of FAS receptor.

These established scenarios allow us to conclude that in the case where primary singlet oxygen that is derived from CAP or PAM, rather than inactivating catalase, would inactivate a SOD molecule or activate a FAS receptor, secondary singlet oxygen generation in an autoamplificatory mode would also be ensured (Fig. 6). Thus, the final outcome of the treatment would be same as that after initial targeting of catalase by singlet oxygen. These homologies of action might well contribute to the overall efficiency of CAP and PAM action.

2. Potential Enhancing Effects of Nitrite

Nitrite is essential as a reaction partner for H_2O_2 to generate peroxynitrite [Fig. 7(A)]. Nitrite has no direct apoptosis-inducing potential up to concentrations of 1 mm. Although (2) and (3) in Fig. 7(A) that lead to singlet oxygen generation through further reactions of peroxynitrite can be counteracted by the efficient reaction of peroxynitrite with CO_2

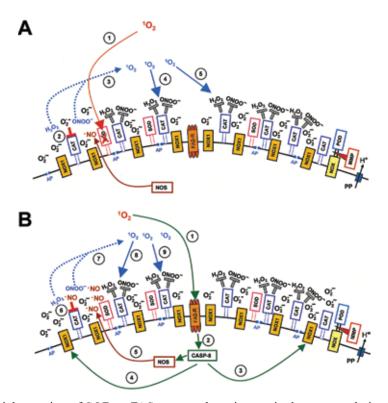


FIG. 6: Initial targeting of SOD or FAS receptor by primary singlet oxygen derived from CAP or PAM. When SOD (A) or FAS receptor (B) are targeted by primary singlet oxygen instead of abundant catalase [as shown in Fig. 4(A)], autoamplification of secondary singlet oxygen generation and catalase inactivation are nevertheless warranted. This is achieved through (1)–(4) in (A) and (1),(7) in (B).

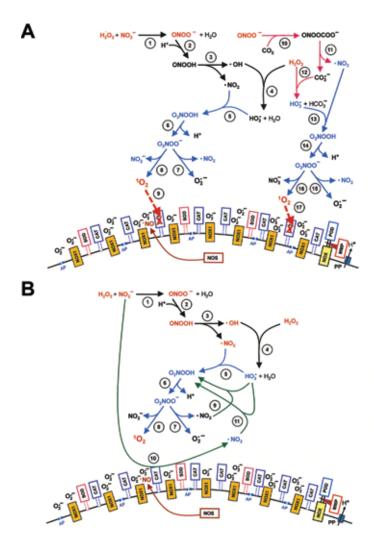


FIG. 7: Plasticity of singlet oxygen generation through interaction between the long-lived species H_2O_2 and nitrite. (A) Role of peroxynitrite. Peroxynitrite is centrally involved in generation of singlet oxygen through the interaction between H_2O_2 and nitrite, following (1)–(9), as shown earlier. This pathway is more likely to occur close to the membrane, due to the presence of proton pumps. As negative side effect, this pathway is minimized by active catalase on tumor cells. Because it is farther away from the cells, peroxynitrite is more likely to interact with CO_2 (10). Decomposition of nitrosoperoxycarboxylate (11) and the reaction between resultant carbonate radicals with H_2O_2 (12) should allow for generation of singlet oxygen through (13)–(16). (B) Role of nitrite and NO_2 . Nitrite (NO_2^-) in PAM or CAP is essential for generation of peroxynitrite (1), whereas the concentration of NO_2 is rate limiting for generation of peroxynitric acid (5). In concert, these reactions lead to formation of singlet oxygen (1)–(8). Even if peroxynitrate (O_2NOO^-) fails to contribute to generation of singlet oxygen due to (7), NO_2 generated through this abortive reaction has a chance to contribute to a new round of peroxynitric acid generation through (9). Even more important for the supply with NO_2 is the oxidation of abundant nitrite by catalase (10).

(10), the overall generation of singlet oxygen are not necessarily reduced, because (11) and (12) lead to generation of hydroperoxyl radicals that are essential for the formation of peroxynitric acid and subsequent generation of singlet oxygen through spontaneous decomposition of peroxynitrate. Thus, the variable reaction potential of peroxynitrite may ensure that the final outcome of singlet oxygen generation is not altered by variable reaction profiles. However, the biological effect of singlet oxygen generated through either pathway is only ensured if singlet oxygen does reach its target structure.

Even if peroxynitrate decomposes into superoxide anions and nitrogen dioxide [(7) in Fig. 7(B)] rather than into singlet oxygen and nitrite [(8) in Fig. 7(B)], this may not necessarily reduce overall singlet oxygen generation. NO₂ derived from (7) may contribute to generation of peroxynitric acid through (9), thus producing a second chance to enhance singlet oxygen generation.

In light of the relative abundance of nitrite in PAM and the potential of catalase to oxidate nitrite to nitrogen dioxide [(10) in Fig. 7(B)], (11) can substantially enhance formation of peroxynitric acid and singlet oxygen. Thus, nitrite in PAM does not seem to be solely necessary for initial generation of peroxynitrite (1) but might also enhance singlet oxygen generation after its oxidation by catalase. The enhancing effects of nitrite and NO₂ most likely contribute to the high efficiency of CAP and PAM. Especially, oxidation of CAP-or PAM-derived nitrite to NO₂ will also contribute to efficient secondary singlet oxygen generation because the otherwise rate-limited NO₂ is then available in relative abundance.

Established reactions between nitrite and catalase are shown in Fig. 8. ¹³⁹ Figure 8(A) summarizes three essential functions of catalase (decomposition of H₂O₂ and peroxynitrite, oxidation of NO) and points to the role of compound I in these reactions. Figure 8(B) shows that nitrite can compete with NO for interactions with compounds I and II. As a result, the local concentration of NO would be enhanced and lead to formation of additional peroxynitrite. Due to the presence of PAM- or CAP-derived H₂O₂ and competition with peroxynitrite for native catalase, the chances for free peroxynitrite survival may thus increase. This may then lead to generation of secondary singlet oxygen through (9)–(14) in Fig. 8.

Alternatively, if the competition between nitrite and NO leads to a sufficiently high NO generation to result in local catalase inhibition [Fig. 9(A)], the resultant free $\rm H_2O_2$ and peroxynitrite may establish generation of singlet oxygen and thus establish autoamplification of singlet oxygen generation and catalase inactivation.

If CAP- or PAM-derived H_2O_2 transforms compound II (formed through the reaction between nitrite and compound I of catalase) into the terminally inactive compound III, free H_2O_2 and peroxynitrite might likewise generate singlet oxygen and trigger autoamplification [Fig. 9(B)]. This pathway is much more likely to occur after nitrite/catalase interaction than after NO/catalase interaction, because the concentration of nitrite is several orders of magnitude higher than that of NO.

D. Apoptosis-Inducing Signaling after CAP and PAM Action on Tumor Cells

Following catalase inactivation by secondary singlet oxygen, NO/peroxynitrite and/or HOCl signaling can be established [Fig. 10(A)]. This occurs analogous to the situation

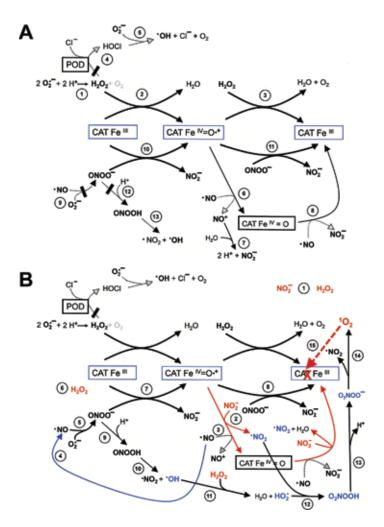


FIG. 8: Reaction between catalase and nitrite. (A) Basic reactions of catalase. Catalase decomposes H_2O_2 in a two-step reaction (1)–(3). This prevents apoptotic induction of tumor cells through the HOCl signaling pathway (4),(5). Compound I of catalase (CATFe^{IV} = $O^{\bullet +}$), the active intermediate obtained after catalase interaction with one molecule of H_2O_2 , oxidates NO in a two-step reaction, with compound II (CATFe^{IV} = O) acting as intermediate (6)–(8). Eventually, formed peroxynitrite (9) is decomposed by catalase in a two-step reaction (10),(11), thus preventing apoptosis-inducing NO/peroxynitrite signaling (12),(13). (B) Interaction between nitrite and catalase. Nitrite contained in PAM or CAP (1) can compete with NO in the interaction with compound I of catalase (2),(3). Due to the abundance of nitrite, this process can be expected to be dominant. Free NO (4) then has a high chance of reacting with superoxide anions to form peroxynitrite (5). Relatively abundant H_2O_2 (6) from PAM or CAP competes with peroxynitrite for interaction with catalase and thus reduces or prevents (7) and (8). This then allows for singlet oxygen generation through (9)–(14), followed by inactivation of additional catalase molecules (15). In this scenario, peroxynitrite formation through interaction between nitrite and H_2O_2 is not required. A detailed description of catalase function and regulation is available elsewhere.¹¹⁴

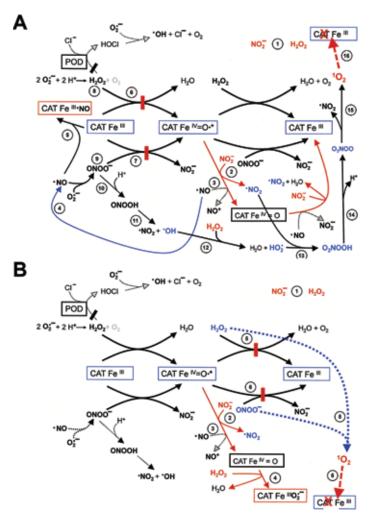


FIG. 9: Alternative modes of nitrite action. (A) NO-mediated inhibition of catalase. Nitrite from CAP or PAM (1) efficiently competes with NO for compound I and oxidates to NO₂ (2),(3). If free NO reaches local concentrations > 0.18 μM, an inactive NO-catalase complex is formed (5).⁷⁴ As a result, H_2O_2 and peroxynitrite are not decomposed at the site of inhibited catalase and generate singlet oxygen through (9)–(15). Singlet oxygen then inactivates further catalase molecules (16). (B) Formation of inactive compound III after nitrite action. Efficient competition of nitrite with NO as well as H_2O_2 for compound I might lead to a transiently high concentration of compound II (CATFe^{IV} = O) that is converted to the terminally inactive compound III (CAT-Fe^{III} O_2 • -). As a result, free H_2O_2 and peroxynitrite generate singlet oxygen according to (9)–(15), as described in (A).

found for other modes of catalase inhibition or inactivation. 75,108,111–113 In the continuous presence of CAP- or PAM-derived H₂O₂, HOCl signaling is favored (Bauer, manuscript in preparation). HOCl and NO/peroxynitrite signaling pathways are finalized by genera-

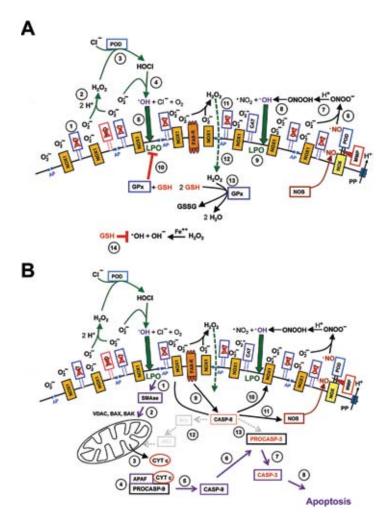


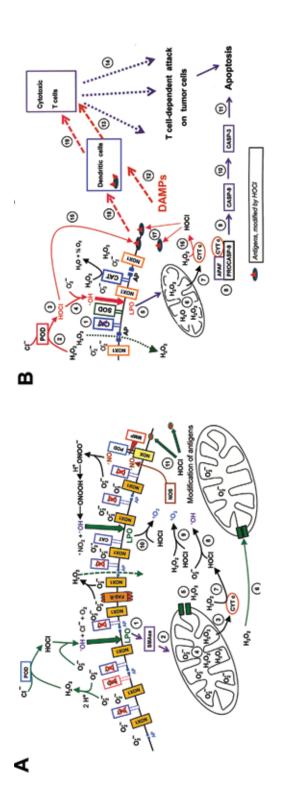
FIG. 10: Intercellular ROS signaling after singlet oxygen—mediated inactivation of tumor cell catalase. (A) Balance between extracellular ROS signaling and intracellular glutathione-mediated protection. After sufficient singlet oxygen—mediated catalase inactivation on the outer surface of tumor cells, the HOCl signaling pathway (1)–(5) and potentially the NO/peroxynitrite signaling pathway (6)–(9) are established. Both pathways are finalized by hydroxyl radical—mediated lipid peroxidation. Initially, glutathione peroxidase and glutathione counteract apoptosis induction by lipid peroxides. However, the lack of protection by inactivated catalase allows strong influx of cell-derived H₂O₂ into cells, leading to efficient depletion of glutathione. This renders the cells vulnerable to apoptosis induction by lipid peroxidation and intracellular hydroxyl radicals. (B) Mitochondrial pathway of apoptosis. Lipid peroxides (1) activate sphingomyelinase (2) and trigger the mitochondrial pathway of apoptosis, mediated by voltage-dependent anion channel (VDAC) and proteins BAK and BAX and characterized by release of cytochrome c (3), formation of apoptosome (4), processing of inactive caspase-9 to active caspase-9 (5), and followed by activation of executing caspase-3 (6)–(8).¹⁴¹ The role of caspase-8 in this scenario is restricted to (9)–(11), as long as FAS receptor is typically expressed at low levels.

tion of hydroxyl radicals that trigger lipid peroxidation. Lipid peroxides can be removed by intracellular glutathione peroxidase and glutathione. This repair reaction initially interferes with hydroxyl radical—mediated apoptosis induction. However, local inactivation of catalase by singlet oxygen allows aquaporin-dependent influx of extracellular H_2O_2 (either derived from CAP or PAM or generated through dismutation of NOX1-generated superoxide anions). Subsequent glutathione peroxidase/glutathione—mediated decomposition of H_2O_2 leads to decreased intracellular glutathione concentration. Therefore, interference of glutathione peroxidase-4/glutathione with lipid peroxidation-dependent triggering of the mitochondrial pathway of apoptosis is gradually abrogated. This central control step explains the findings by Yan et al., who have convincingly shown that aquaporins play a central part during CAP- and PAM-dependent apoptosis induction in malignant cells. Recent experimental evidence supports the concept that these findings extend to other ROS/RNS-mediated apoptosis-inducing principles in general.

Following lipid peroxidation and glutathione depletion, the mitochondrial pathway of apoptosis is activated, ¹⁴¹ summarized in Fig. 10(B). Essential elements are sphingomyelinase; mitochondria; the apoptosome consisting of apoptotic protease-inhibiting factor, cytochrome c, and procaspase-9; and active caspase-9 and -3. Mitochondria involvement in the apoptosis pathway includes release of cytochrome c and decoupling of the mitochondrial respiratory chain. As a result of decoupling, mitochondria involved in apoptosis induction generate high concentrations of superoxide anions that are dismutated to H₂O₂ by mitochondrial SOD [Fig. 11(A)]. H₂O₂ may then reach neighboring intact mitochondria and cause ROS-dependent ROS release through oxidation of the permeability transition pore, followed by uncoupling of the mitochondrial respiratory chain. ¹⁴² In total, this scenario explains the massive increase in ROS (i.e., primarily H₂O₂) in cells that undergo the mitochondrial pathway of apoptosis. ROS increase may promote oxidative effects in the cell population, as suggested by Pletjushkina et al. ¹⁴³

Cytochrome c that is released from mitochondria has recently been found to use H_2O_2 to generate HOCl in a reaction analogous to myeloperoxidase or the perioxidase domain of dual oxidase (Bauer, unpublished result). HOCl that is generated inside cells through this pathway may (1) oxidate tumor cell antigens, thus making them attractive to dendritic cells, 144–149 (2) interact with superoxide anions and generate damaging hydroxyl radicals, 124,129 or (3) react with either H_2O_2 or lipid peroxides and generate singlet oxygen. The biological relevance of these massive secondary reactions has not yet been fully investigated. Detection of mitochondrial singlet oxygen by Bekeschus et al. The may be due to these reactions following the execution of the mitochondrial pathway of apoptosis, although these authors may have missed detecting primary and secondary singlet oxygen on the membrane due to the scavenging function of membrane-associated catalase.

The role of HOCl does not seem to be restricted to apoptosis induction through the HOCl signaling pathway. Due to its high reactivity, HOCl can also very efficiently modify tumor cell antigens and enhance T-cell responses directed to tumor cells. 144-149 It is conceivable that HOCl-dependent antigenic modification may act in concert with



permeability transition pore of neighboring mitochondria and induce ROS-dependent ROS release (6). 142 In total, high concentrations of ture that allows generation of HOCl (7) (Bauer, unpublished result). HOCl may interact with superoxide anions (8), leading to generation of ditional generation of singlet oxygen (10) 151,152 Finally, HOCl may react with cellular proteins and render them more attractive for dendritic cells, the primary cells of immune responses (11). 144-149 (B) Immunogenic cell death. Inactivation of catalase allows for establishment of the lowing lipid peroxidation, activation of sphingomyelinase, release of cytochrome c from mitochondria (1)–(3), and uncoupling of the mitochondrial respiratory chain cause generation of superoxide anions. These dismutate to H,O, both spontaneously and through activity of mitochondrial SOD (4). H,O, can leak out from mitochondria through aquaporins and potentially through VDAC (5). It may act on the ROS are generated through these effects. Recently, cytochrome c has been shown to react with H,O, to form a compound I-analogous struchydroxyl radicals, or with H₂O₂(9), leading to generation of singlet oxygen. ¹⁵⁰ The interaction of HOCl with lipid peroxides may cause admitochondrial pathway of apoptosis (1)–(11). Death-associated molecular patterns (DAMPs) stimulate dendritic cells to provoke a cytotoxic T-cell response (12),(13) that is directed to tumor cells (14). In an analogous mode, HOCl generated through primary HOCl signaling (1) or through cytochrome c-dependent HOCl synthesis (16) modifies tumor cell antigens (17). This seems to generate a DAMP-analogous FIG. 11: Further consequences of the mitochondrial pathway of apoptosis. (A) Secondary mitochondria-dependent ROS generation. Folsignaling pathway, climaxing in a cytotoxic T-cell response (18),(19). 144-149

canonical death-associated molecular patterns that are involved in classical immunogenic cell death that triggers a T-cell response to tumor cells [Fig. 11(B)]. There is general consent that T-cell responses after initial tumor treatment are of central importance to overall outcome of most if not all antitumor therapies presently under investigation. 77,87-91

III. CONCLUSIONS

The selective action of CAP and PAM on tumor cells seems to be based on the interaction between singlet oxygen (derived from CAP or generated through the interaction between H₂O₂ and nitrite in CAP or PAM) and the specific redox biology-related enzyme composition on the surface of tumor cells. Of central and determining importance in this context are membrane-associated catalase and SOD, because they interact with reaction products of membrane-associated NOX and intracellular NOS. FAS receptor also has a modulatory role in this context.

This cooperative system of enzymes and FAS receptor represents a tumor cell–specific biochemical switchboard that can be triggered by singlet oxygen. Catalase and SOD can be inactivated by singlet oxygen, whereas FAS receptor is activated by singlet oxygen. Thus, even a very low concentration of singlet oxygen that is derived from CAP or PAM can establish a strong and sustained biochemical and cellular response through this mechanism. The maintained and autoamplificatory nature of this response is due to the

- (1) establishment of local concentrations of free H₂O₂ and peroxynitrite at the site of inactivated catalase molecules,
- (2) potential of H₂O₂ and peroxynitrite to interact in a complex series of reactions, yielding *de novo* generation of secondary singlet oxygen,
- (3) attack of neighboring active catalase and SOD by the secondary singlet oxygen.

After inactivation of a sufficiently high portion of protective catalase, intercellular ROS/RNS-mediated apoptosis-inducing signaling is established and causes elimination of malignant cells. Because this biochemical switchboard is unavailable to nonmalignant cells, selectivity of CAP and PAM action for tumor cells is ensured as long as the dose of CAP and PAM is in a range that does not allow nonselective apoptosis induction in nonmalignant cells by hydrogen peroxide or peroxynitrite.

These autoamplificatory processes that are activated by singlet oxygen on tumor cells use, in a sustained mode, the same ROS/RNS that also act in CAP and PAM and show a partial overlap to intercellular ROS/RNS-dependent apoptosis-inducing signaling. Essential compounds for the autoamplificatory mechanism are superoxide anions, nitric oxide, nitrogen dioxide, peroxynitrite (ONOO⁻), peroxynitrous acid (ONOOH), hydrogen peroxide, hydroxyl radicals, hydroperoxyl radicals, peroxynitrate (O₂NOO⁻), peroxynitric acid (O₂NOOH), and singlet oxygen. Due to involvement of enzymes and

proton pumps on the tumor cell membrane, this ROS/RNS-dependent process is localized to the membrane. This ensures high efficiency selectivity of these processes.

Catalase inactivation of secondary singlet oxygen far exceeds inactivation of catalase by primary, CAP-, and PAM-generated singlet oxygen. Catalase inactivation allows an essential aquaporin-mediated influx of tumor cell–derived H_2O_2 into the targeted cells, causing depletion of intracellular glutathione. This results in the abrogation of glutathione peroxidase 4/glutathione inhibitory effects to apoptosis-inducing signaling by lipid peroxides. Hydroxyl radicals induce lipid peroxidation that is generated by intercellular ROS/RNS signaling.

The subsequent induction of the apoptosis mitochondrial pathway results in activation of caspase-9 and -3. In parallel, the release of cytochrome c in apoptotic cells increases concentrations of superoxide anions and H_2O_2 , generated by the uncoupled respiratory chain. The potential contribution of these ROS, arising late in the apoptotic process of targeted cells, is considered to contribute to the spread of the apoptotic response in the cell population.¹⁴³ So far, its potential biological relevance seems to be underestimated.

These considerations demonstrate the multiple roles of H_2O_2 for the chemical biology of CAP and PAM action, starting from an involvement in singlet oxygen formation, maintenance and modulation of HOCl signaling, to modulation of apoptotic pathways. These findings add to multiple other roles of H_2O_2 that have been pointed out earlier by Sies. ^{153,154} The strong ROS/RNS-driven autoamplificatory network induced by CAP and PAM has inherent potential for the following:

- synergistic interaction between parallel inactivation of membrane-associated catalase and SOD, because both enzymes are inactivated by singlet oxygen and biochemically interactive,
- enhanced effect of singlet oxygen—mediated activated FAS receptor activation, with an impact on enhancement of superoxide anion and NO production, thus enhancing generation of secondary singlet oxygen,
- oxidation of nitrite to nitrogen dioxide, which is a rate-limiting reaction partner during the generation of secondary singlet oxygen,
- interaction of nitrite with the catalase cycle, potentially slowing enzyme activity
- competition between nitrite and NO for the reaction with compound I of catalase, including potential local increase of NO and subsequent inhibition of catalase.

These potential synergistic interactions between ROS- and RNS-dependent processes of CAP and PAM action are the basis for an additional and essential strong immunological process, termed immunogenic cell death, that is characterized by the release of death-associated molecular patterns (DAMPs). Therefore, the chain of biochemical and cellular events initiated by CAP- or PAM-derived singlet oxygen and amplified by sustained generation of cell-derived secondary singlet oxygen leads to massive finalization of antitumor action through the immune system. HOCl that is primarily involved

in intercellular apoptosis-inducing signaling after the inactivation of catalase may also contribute to immunogenic cell death through modification of tumor cell antigens.

IV. EPILOGUE

We established the novel model of Bauer and Graves^{20,21} in a deductive approach, in which established knowledge of the composition and action of CAP and PAM was connected to experimentally obtained results involving the differential action of defined ROS/RNS on malignant and nonmalignant cells.

This model was confirmed in a bottom-up experimental approach using long-lasting compounds of PAM. A recent experimental approach with CAP and PAM generated by a plasma generator confirmed the results obtained by the preceding model experiments.

ACKNOWLEDGMENTS

We gratefully acknowledge the intellectual support of many colleagues during the 2016, 2017, and 2018 International Workshops on Plasma for Cancer Treatment, 7th International Conference on Plasma Medicine, and 2018 Gordon Research Conference on Plasma Processing Science, which were of central importance to this work. We thank J. Brandel (Freiburg, Germany) for technical assistance and acknowledge the support of Hans-Sauer-Stiftung and The European Cooperation in Science and Technology (COST) Action Grant No. CM0603.

REFERENCES

- 1. Machala Z, Janda M, Hensel K, Jedlovsky I, Lestinska L, Foltin V, Martisovits V, Morvova M. Emission spectroscopy of atmospheric pressure plasmas for bio-medical and environmental applications. J Mol Spectroscopy. 2007;243:194–201.
- 2. Stoffels E, Sakiyama Y, Graves DB. Cold atmospheric plasma: Charged species and their interactions with cells and tissues. IEEE Trans Plasma Sci. 2008;36:1441–57.
- 3. Fridman G, Friedman G, Gutsol A, Shekhter AB, Vasilets VN, Fridman A. Applied plasma medicine. Plasma Proc Polym. 2008;5:503–33.
- Attri P, Arora B, Choi EH. Utility of plasma: A new road from physics to chemistry. RSC Adv. 2013;3:12540-67.
- Graves DB. The emerging role of reactive oxygen and nitrogen species in redox biology and some implications for plasma applications to medicine and biology. J Phys D Appl Phys. 2012;45:263001.
- 6. Graves DB. Mechanisms of plasma medicine: Coupling plasma physics, biochemistry, and biology. IEEE Trans Radiat Plasma Med Sci. 2017;1:281–92.
- 7. Laroussi M. Low-temperature plasma for medicine. IEEE Trans Plasma Sci. 2009;37:714–15.
- 8. Laroussi M. From killing bacteria to destroying cancer cells: 20 years of plasma medicine. Plasma Proc Polym. 2014;11:1138–41.
- 9. Laroussi M. Low-temperature plasma jet for biomedical applications: A review. IEEE Trans Plasma Sci. 2015;43:703–12.
- Laroussi M, Lu X, Keidar M. Perspective: The physics, diagnostics, and applications of atmospheric pressure low temperature plasma sources used in plasma medicine. J Appl Phys. 2017;122:020901.

- Lu X, Naidis GV, Laroussi M, Reuter S, Graves DB, Ostrikov K. Reactive species in non-equilibrium atmospheric-pressure plasmas: Generation, transport, and biological effects. Phys Rep Phys Lett. 2016;630:1–84.
- 12. Graves DB. Reactive species from cold atmospheric plasma: Implications for cancer therapy. Plasma Proc Polym. 2014;11:1120–7.
- Graves DB. Oxy-nitroso shielding burst model of cold atmospheric plasma therapeutics. Clin Plasma Med. 2014;2:38–49.
- Jablonowski H, Bussiahn R, Hammer MU, Weltman K-D, von Woedtke T, Reuter S. Impact of plasma jet vacuum ultraviolet radiation on reactive species generation in bio-relevant liquids. Phys Plasmas. 2015;22:122008.
- Sousa JS, Niemi K, Cox LJ, Algwari QT, Gans T, O'Connell D. Cold atmospheric pressure plasma jets as sources of singlet delta oxygen for biomedical applications. J Appl Phys. 2011;109:123302.
- Machala Z, Tarabova B, Hensel K, Spetlikova E, Sikurova L, Lukes P. Formation of ROS and RNS in water electro-sprayed through transient spark discharge in air and their bactericidal effects. Plasma Proc Polym. 2013;10:649–59.
- Wende K, Williams P, Dalluge J, Van Gaens W, Akoubakr H, Bischof J, von Woedtke T, Goyal SM, Weltmann K-D, Bogaerts A, Masur K, Bruggeman PJ. Identification of biologically active liquid chemistry induced by nonthermal atmospheric pressure plasma jet. Biointerphases. 2015;10:029518.
- 18. Schmidt-Bleker A, Bansemer R, Reuter S, Weltmann K-D. How to produce a NO*x* instead of O*x*-based chemistry with a cold atmospheric plasma jet. Plasma Proc Polym. 2016;13:1120–7.
- Gianela M, Reuter S, Aguila AL, Ritchie GAD, van Helden JPH. Detection of HO₂ in an atmospheric pressure plasma jet using optical feedback cavity-enhanced absorption spectroscopy. New J Phys. 2016;18:113027.
- Bauer G, Graves DB. Mechanisms of selective antitumor action of cold atmospheric plasma-derived reactive oxygen and nitrogen species. Plasma Proc Polym. 2016;13:1157–78.
- 21. Bauer G. Signal amplification by tumor cells: Clue to the understanding of the antitumor effects of cold atmospheric plasma and plasma-activated medium. IEEE Trans Radiat Plasma Med Sci. 2018;2:87–98.
- 22. Yan A, Talbot N, Nourmokammadi X, Cheng J, Canady J, Sherman JH, Keidar M. Principles of using cold atmospheric plasma stimulated media for cancer treatment. Sci Rep. 2015;5:1833901–17.
- 23. Kurake N, Tanaka H, Ishikawa K, Kondo T, Sekine M, Nakamura K, Kajiyama H, Kikkawa F, Mizuno M, Hori M. Cell survival of glioblastoma grown in medium containing hydrogen peroxide and/or nitrite, or in plasma-activated medium. Arch Biochem Biophys. 2016;605:102–8.
- 24. Girard P-M, Arbabian A, Fleury M, Bauville G, PuechV, Dutreix M, Sousa JS. Synergistic effect of H₂O₂ and NO₂⁻ in cell death induced by cold atmospheric He plasma. Sci Rep. 2016;6:29098.
- Uchida G, Nakajima A, Takenaka K, Kawasaki T, Koja K, Shiratani M, Setsuhara Y. Effects of nonthermal plasma jet irradiation on the selective production of H₂O₂ and NO₂⁻ in liquid water. J Appl Phys. 2016;120:201102.
- Dobrynin D, Fridman G, Fridman G, Fridman A. Physical and biological mechanisms of direct plasma interaction with living tissue. New J Phys. 2009;11:115020.
- Vandamme M, Pesnel S, Barbosa E, Dozias S, Sobilo J, Lerondel S, Le Pape A, Pouvesle J-M. Antitumor effect of plasma treatment on U87 glioma xenografts: Preliminary results. Plasma Proc Polym. 2010;7:264–73.
- 28. Von Woedtke T, Metelmann H-R, Weltmann K-D. Clinical plasma medicine: State and perspectives of in vivo application of cold atmospheric plasma. Contrib Plasma Phys. 2014;54:104–17.
- 29. Keidar M. Plasma for cancer treatment. Plasma Sources Sci Technol. 2015;24:033001.
- Yan DY, Sherman JH, Keidar M. Cold atmospheric plasma, a novel promising anti-cancer treatment modality. Oncotarget. 2017;8:15977–95.

 Metelmann H-R, Nedrelow DS, Seebauer C, Schuster M, von Woedtke T, Weltmann K-D, Kindler S, Metelmann PH, Finkelstein SE, Von Hoff DD. Head and neck cancer treatment and physical plasma. Clin Plasma Med. 2015;3:17–23.

- 32. Metelmann H-R, Seebauer C, Miller V, Fridman A, Bauer G, Graves DB, Pouvesle J-M, Rutkowski R, Schuster M, Bekeschus S, Wende K, Masur K, Hasse S, Gerling T, Hori M, Tanaka H, Choi EH, Weltmann K-D, Metelmann PH, von Hoff DD, von Woedtke T. Clinical experience with cold plasma in the treatment of locally advanced head and neck cancer. Clin Plasma Med. 2018;9:6–13.
- Metelmann HR, Seebauer C, Rutkowski R, Schuster M, Bekeschus S, Metelmann P. Treating cancer with cold physical plasma: On the way to evidence-based medicine. Contrib Plasma Phys. 2018;58:415–9.
- 34. Schuster M, Seebauer C, Rutkowski R, Hauschild A, Podmelle F, Metelmann C, Metelmann B, Von Woedtke T, Hasse S, Weltmann K-D, Metelmann H-R. Visible tumor surface response to physical plasma and apoptotic cell kill in head and neck cancer. J Craniomaxillofac Surg. 2016;44:1445–52.
- Weltmann KD, von Woedtke T. Plasma medicine—Current state of research and medical application. Plasma Phys Contr Fusion. 2017;59(1):014
- 36. Machala Z, Chladekova L, Pelach M. Plasma agents in bio-decontamination by DC discharges in atmospheric air. J Phys D Appl Phys. 2010;43:222001.
- 37. Wu H, Sun P, Feng H, Zhou H, Wang R, Liang Y, Lu J, Zhu W, Zhang J, Fang J. Reactive oxygen species in a nonthermal plasma microjet and water system: Generation, conversion and contributions to bacteria inactivation—An analysis by electron spin resonance spectroscopy. Plasma Proc Polym. 2012;9:417–24.
- Traylor MJ, Pavlovich MJ, Karim S, Hait P, Sakiyama Y, Clark DS, Graves DB. Long-term antibacterial efficacy of air plasma-activated water. J Phys D Appl Phys. 2011;44:472001.
- Aboubakr A, Gangal U, Youssef MM, Goyal SM, Bruggeman PJ. Inactivation of virus in solution by cold atmospheric pressure plasma: Identification of chemical inactivation pathways. J Phys D Appl Phys. 2016;48:204001.
- Isbary G, Morfill G, Schmidt HU, Georgi M, Ramrath K, Heinlein J, Karrer S, Landthaler M, Shimizu T, Steffes B, Bunk W, Monetti R, Zimmermann JL, Pompl R, Stolz W. A first prospective randomized controlled trial to decrease bacterial load using cold atmospheric argon plasma on chronic wounds in patients. Br J Dermatol. 2010;163:78–82.
- 41. Arndt S, Unger P, Wacker E, Shimizu T, Heinlin J, Li J-F, Thomas HM, Morfill GE, Zimmermann JL, Bosserhoff A-K, Karrer S. Cold atmospheric plasma (CAP) changes gene expression of key molecules of the wound healing machinery and improves wound healing in vitro and in vivo. PLOS ONE. 2013;8:e79325.
- 42. Hasse S, Tran D, Hahn O, Kindler S, Metelmann H-R, von Woedtke T, Masur K. Induction of proliferation of basal epidermal keratinocytes by cold atmospheric-pressure plasma. Clin Exp Dermatol. 2016;41:202–9.
- 43. Schmidt A, Bekeschus S, Wende K, Vollmar B, von Woedtke T. A cold plasma jet accelerates wound healing in a murine model of full-thickness skin wounds. Exp Dermatol. 2017;26:156–62.
- 44. Heinlin J, Isbary G, Stolz W, Morfill G, Landthaler M, Shimizu T, Steffes B, Nosenko T, Zimmermann JL, Karrer S. Plasma applications in medicine with a special focus on dermatology. J Eur Acad Dermatol Venereol. 2011;25(1):1–11.
- Friedman PC, Miller V, Fridman G, Lin A, Fridman A. Successful treatment of actinic keratosis using nonthermal atmospheric pressure plasma: A case series. J Am Acad Dermatol. 2017;76: 340-50.
- 46. Wirtz M, Stoffels I, Dissemond J, Schadendorf D, Roesch A. Actinic keratoses treated with cold atmospheric plasma. J Eur Acad Dermatol Venereol. 2018;32:e37–9.

- Keidar M, Walk R, Shashurin A, Srinivasan P, Sandler P, Sandler A, Dasgupta S, Ravi R, Guerrero-Preston R, Trink B. Cold plasma selectivity and the possibility of a paradigm shift in cancer therapy. Br J Cancer. 2011;105:1295–301.
- Keidar M, Shashurin A, Volotskova O, Stepp MA, Srinivasan P, Sandler A, Trink B. Cold atmospheric plasma in cancer therapy. Phys Plasma. 2013;20:057101.
- Ishaq M, Evans M, Ostrikov K. Effect of atmospheric gas plasmas on cancer cell signaling. Int J Cancer. 2013;134:1517–28.
- Barekzi N, Laroussi M. Effects of low temperature plasmas on cancer cells. Plasma Proc Polym. 2013;10:1039–50.
- 51. Schlegel J, Köritzer J, Boxhammer V. Plasma in cancer treatment. Clin Plasma Med. 2013;1:2–7.
- 52. Tanaka H, Mizuno M, Ishikawa K, Takeda K, Nakamura K, Utsumi F, Kajiyama H, Kano H, Okazaki Y, Toyokuni S, Maruyama S, Kikkawa F, Hori M. Plasma medical science for cancer therapy: Toward cancer therapy using nonthermal atmospheric pressure plasma. IEEE Trans Plasma Sci. 2013;42:3760–4.
- Ratovitski EA, Heng X, Yan D, Sherman JH, Canady J, Trink B, Keidar M. Anti-cancer therapies of the 21st century: Novel approach to treat human cancers using cold atmospheric plasma. Plasma Proc Polym. 2014;11:1128–37.
- 54. Babington P, Rajjoub K, Canady J, Siu A, Keidar M, Sherman JH. Use of cold atmospheric plasma in the treatment of cancer. Biointerphases. 2015;10:029403.
- Gay-Mimbrera J, Garcia MC, Tejera BI, Rodero-Serrano A, Garcia-Nieto AV, Ruano J. Clinical and biological principles of cold atmospheric plasma application in skin cancer. Adv Ther. 2016;33:894

 –909.
- 56. Bauer G, Graves DB, Schuster M, Metelmann H-R. Side effect management. In: Metelmann H-R, von Woedtke T, Weltmann K-D, editors. Comprehensive clinical plasma medicine. Berlin: Springer; 2pp. 301–18.
- 57. Kalghati S, Kelly C, Cerchar E, Azizkhan-Clifford J. Selectivity of non-thermal atmospheric-pressure microsecond-pulsed dielectric barrier discharge plasma induced apoptosis in tumor cells over healthy cells. Plasma Med. 2011;1:249–63.
- 58. Tanaka H, Ishikawa K, Nakamura K, Kajiyama H, Komo H, Kikkawa T, Hori M. Plasma-activated medium selectively kills glioblastoma brain tumor cells by down-regulating a survival signaling molecule, AKT kinase. Plasma Med. 2011;1:265–77.
- Zucker SN, Zirnheld J, Bagati A, DiSanto TM, Des Soye B, Wawrzyniak JA, Eternadi K, Nikiforov M, Berezney R. Preferential induction of apoptotic cell death in melanoma cells as compared with normal keratinocytes using a non-thermal plasma torch. Cancer Biol Ther. 2012;13:1299–306.
- 60. Wang M, Holmes B, Cheng X, Zhu W, Keidar M, Zhang LG. Cold atmospheric plasma for selectively ablating metastatic breast cancer cells. PLOS ONE. 2013;8:e73741.
- Guerrero-Preston R, Ogawa T, Uemura M, Shumulinsky G, Valle BL, Pirini F, Ravi R, Sidransky D, Keidar M, Trink B. Cold atmospheric plasma treatment selectively targets head and neck squamous cell carcinoma cells. Int J Mol Med. 2014;34:941–6.
- 62. Kaushik N, Kumar N, Kim CH, Kaushik N, Choi EH. Dielectric barrier discharge plasma efficiently delivers an apoptotic response in human monocytic lymphoma. Plasma Proc Polym. 2014;11:1175–87.
- Utsumi F, Kajiyama H, Nakamura K, Tanaka H, Hori M, Kikkawa F. Selective cytotoxicity of indirect nonequilibrium atmospheric pressure plasma against ovarian clear-cell carcinoma. Springerplus. 2014;3:398.
- 64. Siu A, Volotskova O, Cheng X, Khaisa SS, Bian K, Murad F, Keidar M, Sherman JH. Differential effects of cold atmospheric plasma in the treatment of malignant glioma. PLOS ONE. 2015;10:e0126313.
- 65. Kim SJ, Chung TH. Cold atmospheric plasma jet-generated RONS and their selective effects on normal and carcinoma cells. Sci Rep. 2016;6:20332.
- 66. Duan J, Lu X, He G. The selective effect of plasma-activated medium in an in vitro co-culture of liver cancer and normal cells. J Appl Phys. 2017;121:013302.

Canal C, Fontelo R, Hamouda I, Guillem-Marti J, Cvelbar U, Ginebra M-P. Plasma-induced selectivity in bone cancer cell death. Free Radic Biol Med. 2017;110:72–80.

- 68. Liedke KR, Bekeschus S, Kaeding A, Hackbarth C, Kuehn JP, Heidecke CD, von Bernstorff W, von Woedtke T, Partecke LI. Non-thermal plasma-treated solution demonstrates antitumor activity against pancreatic cancer cells in vitro and in vivo. Sci Rep. 2017;7:8319.
- 69. Wende K, Straßenburg S, Haertel B, Harms M, Holtz S, Barton A, Masur K, von Woedtke T, Lindequist U. Atmospheric pressure plasma jet treatment evokes transient oxidative stress in HaCaT keratinocytes and influences cell physiology. Cell Biol Int. 2014;38:412–25.
- Hirst AM, Simms MS, Mann VM, Maitland NJ, O'Connell D, Frame FM. Low temperature plasma treatment induces DNA damage leading to necrotic cell death in primary prostate epithelial cells. Br J Cancer. 2015;112:1536–45.
- 71. Bekeschus S, Masur K, Kolata J, Wende K, Schmidt A, Bundscherer L, Barton A, Kramer A, Bröker B, Weltmann K-D. Human mononuclear cell survival and proliferation is modulated by cold atmospheric plasma jet. Plasma Proc Polym. 2013;10:706–13.
- 72. Bekeschus S, Iseni S, Reuter S, Masur K, Weltmann K-D. Nitrogen shielding of an argon plasma jet and its effects on human immune cells. IEEE Trans Plasma Sci. 2015;43:776–81.
- 73. Bundscherer L, Bekeschus S, Tresp H, Hasse S, Reuter S, Weltmann K-D, Lindequist U, Masur K. Viability of human blood leucocytes compared with their respective cell lines after plasma treatment. Plasma Med. 2013;3:71–80.
- Vandamme M, Robert E, Lerondel S, Sarron V, Ries D, Dozias S, Sobilo J, Gosset D, Kieda C, Legrain B, Pouvesle J-M, Le Pape A. ROS implication in a new antitumor strategy based on nonthermal plasma. Int J Cancer. 2012;130:2185–94.
- 75. Bauer G. Increasing the endogenous NO level causes catalase inactivation and reactivation of intercellular apoptosis signaling specifically in tumor cells. Redox Biol. 2015;6:353–71.
- 76. Bauer G. The antitumor effect of singlet oxygen. Anticancer Res. 2016;36:5649–64.
- 77. Garg AD, Agostinos P. ER stress, autophagy and immunogenic cell death in photodynamic therapy-induced anti-cancer immune responses. Photochem Photobiol Sci. 2014;13:474–87.
- 78. Lin A, Truong B, Pappas A, Kirifides L, Oubarri A, Chen S, Lin S, Dobrynin D, Fridman G, Fridman A, Sang N, Miller V. Uniform nanosecond pulsed dielectric barrier discharge plasma enhances antitumor effects by induction of immunogenic cell death in tumors and stimulation of macrophages. Plasma Proc Polym. 2015;12:1392–9.
- Lin A, Truong B, Patel S, Kaushik N, Choi EH, Fridman G, Fridman A, Miller V. Nanosecond-pulsed DBD plasma-generated reactive oxygen species trigger immunogenic cell death in A549 lung carcinoma cells through intracellular oxidative stress. Int J Mol Sci. 2017;18:966.
- Lin A, Truong B, Fridman G, Fridman A, Miller V. Immune cells enhance selectivity of nanosecondpulsed DBD plasma against tumor cells. Plasma Med. 2017;7:85–96.
- 81. Lin AG, Xiang B, Merlino DJ, Baybutt TR, Sahu J, Fridman A, Snook AE, Miller V. Non-thermal plasma induces immunogenic cell death in vivo in murine CT26 colorectal tumors. Oncoimmunology. 2018;7:e148978.
- 82. Miller V, Lin A, Fridman A. Why target immune cells for plasma treatment of cancer? Plasma Chem Plasma Proc. 2016;36:259–68.
- 83. Mizuno K, Yonetamari Y, Shirakawa Y, Akiyama T, Ono R. Anti-tumor immune response induced by nanosecond pulsed streamer discharge in mice. J Phys D Appl Phys. 2017;50:12LT01.
- 84. Bekeschus S, Mueller A, Miller V, Gaipl U, Weltmann K-D. Physical plasma elicits immunogenic cancer cell death and mitochondrial singlet oxygen. IEEE Trans Rad Plasma Med Sci. 2018;2(2): 138–46.
- 85. Bekeschus S, Clemen R, Metelmann H-R. Potentiating anti-tumor immunity with physical plasma. Clin Plasma Med. 2018;12:17–22.

- 86. Kaushik NK, Kaushik N, Min B, K. Choi KH, Hong YJ, Miller V, Fridman A, Choi EH. Cytotoxic macrophage-released tumour necrosis factor-α (TNF-α) as a killing mechanism for cancer cell death after cold plasma activation. J Phys D Appl Phys. 2018;49:084001.
- 87. Apetoh L, Tesnier A, Ghiringhelli F, Kroemer G, Zitvogel L. Interactions between dying tumor cells and the innate immune system determine the efficiency of conventional antitumor therapy. Cancer Res. 2008;68:4026–30.
- Green DR, Ferguson T, Zitvogel L, Kroemer G. Immunogenic and tolerogenic cell death. Nat Rev Immunol. 2009;9:353–63.
- Krysko DV, Garg AD, Kaczmarek A, Krysko O, Agostinis P, Vandenabeele P. Immunogenic cell death and DAMPs in cancer therapy. Nat Rev Cancer. 2012;12:860–75.
- Kroemer G, Galluzzi L, Kepp O, Zitvogel L. Immunogenic cell death in cancer therapy. Ann Rev Immunol. 2013;31:51–72.
- 91. Candeias SM, Gaipl US. The immune system in cancer prevention, development and therapy. Anticancer Agents Med Chem. 2016;16:101–7.
- Utsumi F, Kajiyama H, Nakamura K, Tanaka H, Mizuno M, Ishikawa K, Kondo H, Kano H, Hori M, Kikkawa F. Effect of indirect nonequilibrium atmospheric pressure plasma on anti-proliferative activity against chronic chemoresistant ovarian cancer cells in vitro and in vivo. PLOS ONE. 2013;8:e81576.
- 93. Yan D, Sherman JH, Cheng X, Ratovitski E, Canady J, Keidar M. Controlling plasma stimulated media in cancer treatment application. Appl Phys Lett. 2014;105:22410101–4.
- 94. Adachi T, Tanaka H, Nonomura S, Hara H, Kondo S-I, Hori M. Plasma-activated medium induces A459 cell injury via a spiral apoptotic cascade involving the mitochondrial-nuclear network. Free Radic Biol Med. 2015;79:28–44.
- Mohades S, Laroussi M, Sears J, Barekzi N, Razavi H. Evaluation of the effects of a plasma-activated medium on cancer cells. Phys Plasmas. 2015;22:122001.
- 96. Kumar N, Park JH, Jeon SN, Park BS, Chori EH, Attri P. The action of microsecond-pulsed plasma-activated media on the inactivation of human lung cancer cells. J Phys D Appl Phys. 2016;49:11540101–9.
- 97. Yan D, Nourmohammadi N, Bian K, Murad F, Sherman JH, Keidar M. Stabilizing the cold plasmastimulated medium by regulating medium's composition. Sci Rep. 2016;6:2602.
- 98. Koensgen D, Besic I, Gumbel D, Kaul A, Weiss M, Diesing K, Kramer A, Bekeschus S, Mustea A, Stope MB. Cold atmospheric plasma (CAP) and CAP-stimulated cell culture media suppress ovarian cancer cell growth—A putative treatment option in ovarian cancer therapy. Anticancer Res. 2017;37:6739–44.
- Szili EJ, Bradley JW, Short RD. A "tissue model" to study the plasma delivery of reactive oxygen species. J Phys D Appl Phys. 2014;47:152002.
- 100. Szili EJ, Oh JS, Hong SH, Hatta A, Short RD. Probing the transport of plasma-generated RONS in an agarose target as surrogate for real tissue: Dependency on time, distance and material composition. J Phys D Appl Phys. 2015;48:202001.
- 101. Szili EJ, Hong S-H, Oh J-S, Gaur N, Short RD. Tracking the penetration of plasma reactive species in tissue models. Trends Biotechnol. 2018;36:594–602.
- 102. Szili EJ, Oh JS, Fukuhara H, Bhatia R, Gaur N, Nguyen CK, Hong SH, Ito S, Ogawa K, Kawada C, Shuin T, Tsuda M, Furikata M, Kurabayashi A, Furuta H, Ito M, Inoue K, Hatta A, Short RD. Modelling the helium plasma jet delivery of reactive species into a 3D cancer tumor. Plasma Sources Technol. 2018;27:014001.
- 103. Nie L, Yang Y, Duan J, Sun F, Lu X, He G. Effect of tissue thickness and liquid composition on the penetration of long-lifetime reactive oxygen and nitrogen species (RONS) generated by plasma jet. J Phys D Appl Phys. 2018;51:345204.

104. Ivanovas B, Bauer G. Selective and nonselective apoptosis induction in transformed and nontransformed fibroblasts by exogenous reactive oxygen and nitrogen species. Anticancer Res. 2002;22: 841–56.

- 105. Böhm B, Heinzelmann S, Motz M, Bauer G. Extracellular localization of catalase is associated with the transformed state of malignant cells. Biol Chem. 2015;396:1339–56.
- Bechtel W, Bauer G. Catalase protects tumor cells against apoptosis induction by intercellular ROS signaling. Anticancer Res. 2009;29:4541–57.
- Bechtel W, Bauer G. Modulation of intercellular ROS signaling of human tumor cells. Anticancer Res. 2009;29:4559–70.
- Heinzelmann S, Bauer G. Multiple protective functions of catalase against intercellular apoptosisinducing ROS signaling of human tumor cells. Biol Chem. 2010;391:675–93.
- 109. Bauer G, Zarkovic N. Revealing mechanisms of selective, concentration-dependent potentials of 4-hydroxy-2-nonenal to induce apoptosis in cancer cells through inactivation of membrane-associated catalase. Free Radic Biol Med. 2015;81:128–44.
- Bauer G, Motz M. The antitumor effect of single-domain antibodies directed towards membraneassociated catalase and superoxide dismutase. Anticancer Res. 2016;36:5945

 –56.
- 111. Bauer G. Tumor cell protective catalase as a novel target for rational therapeutic approaches based on specific intercellular ROS signaling. Anticancer Res. 2012;32:2599–24.
- 112. Bauer G. Targeting extracellular ROS signaling of tumor cells. Anticancer Res. 2014;34:1467–82.
- 113. Bauer G. SiRNA-based analysis of the abrogation of the protective function of membrane-associated catalase of tumor cells. Anticancer Res. 2017;37:567–82.
- Bauer G. Targeting the protective catalase of tumor cells with cold atmospheric plasma-treated medium (PAM). Anticancer Agents Med Chem. 2018;18:784

 –804.
- 115. Riethmüller M, Burger N, Bauer G. Singlet oxygen treatment of tumor cells triggers extracellular singlet oxygen generation, catalase inactivation and reactivation of intercellular apoptosis-inducing signaling. Redox Biol. 2015;6:157–68.
- Scheit K, Bauer G. Direct and indirect inactivation of tumor cell protective catalase by salicylic acid and anthocyanidins reactivates intercellular ROS signaling and allows for synergistic effects. Carcinogenesis. 2015;36:400–11.
- Bauer G. Autoamplificatory singlet oxygen generation sensitizes tumor cells for intercellular apoptosis-inducing signaling. Mech Ageing Dev. 2018;172:59–77.
- Yan DY, Talbot A, Nourmohammadi N, Sherman JH, Cheng XQ, Keidar M. Toward understanding the selective anticancer capacity of cold atmospheric plasma—A model based on aquaporins. Biointerphases. 2015;10:040801.
- Yan D, Xiao H, Zhu W, Nourmohammadi N, Zhang LG, Bian K, Keidar M. The role of aquaporins in the anti-glioblastoma capacity of the cold plasma-stimulated medium. J Phys D Appl Phys. 2017;50:055401.
- Irani Y, Xia J, Zweier L, Sollott SJ, Der CJ, Fearon ER, Sundaresan M, Finkel T, Goldschmidt-Clermont PJ. Mitogenic signalling by oxidants in Ras-transformed fibroblasts. Science. 1997;275:1649–52.
- 121. Herdener M, Heigold S, Saran M, Bauer G. Target cell-derived superoxide anions cause efficiency and selectivity of intercellular induction of apoptosis. Free Radic Biol Med. 2000;29:1260–71.
- Heigold S, Sers C, Bechtel W, Ivanovas B, Schäfer R, Bauer G. Nitric oxide mediates apoptosis induction selectively in transformed fibroblasts compared to nontransformed fibroblasts. Carcinogenesis. 2002;23:929

 –41.
- 123. Bauer G. Nitric oxide contributes to selective apoptosis induction in malignant cells through multiple reaction steps. Crit Rev Oncogen. 2016;21:365–98.
- 124. Bauer G. HOCl and the control of oncogenesis. J Inorgan Biochem. 2018;179:10-23.
- 125. Kono Y, Fridovich I. Superoxide radical inhibits catalase. J Biol Chem. 1982;257:5751-4.
- Shimizu N, Kobayashi K, Hayashi K. The reaction of superoxide radical with catalase. Mechanism of the inhibition of catalase by superoxide radical. J Biol Chem. 1984;259:4414–18.

- Gebicka L, Metodiewa D, Gebicki JL. Pulse radiolysis of catalase in solution. I. Reactions of O₂
 with catalase and its compound. Int J Rad Biol. 1989;55:45–50.
- Lardinois OM. Reactions of bovine liver catalase with superoxide radicals and hydrogen peroxide.
 Free Rad Res. 1995;22:251–74.
- 129. Bauer G. HOCl-dependent singlet oxygen and hydroxyl radical generation modulate and induce apoptosis of malignant cells. Anticancer Res. 2013;33:3589–602.
- Escobar JA, Rubio A, Lissi EA. SOD and catalase inactivation by singlet oxygen and peroxyl radicals. Free Rad Biol Med. 1996;20:285–90.
- Kim YK, Kwon OJ, Park J-W. Inactivation of catalase and superoxide dismutase by singlet oxygen derived from photoactivated dye. Biochimie. 2001;83:437

 –44.
- 132. Jablonowski H, von Woedtke T. Research on plasma medicine-relevant plasma-liquid interaction: What happened in the past five years? Clin Plasma Med. 2015;3:42–52.
- Whiteside C, Hassan HM. Role of oxyradicals in the inactivation of catalase by ozone. Free Rad Biol Med. 1988;5:305–12.
- Lee Y-K, Kim SMK, Hand S. Ozone-induced inactivation of antioxidant enzymes. Biochimie. 2003; 85:947–52.
- 135. Zhuang S, Demir JT, Kochevar IE. Protein kinase C inhibits singlet oxygen-induced apoptosis by decreasing caspase-8 activation. Oncogene. 2001;20:6764–76.
- Suzuki Y, Ono Y, Hirabayashi Y. Rapid and specific reactive oxygen species generation via NADPH oxidase activation during FAS-mediated apoptosis. FEBS Lett. 1998;425:209–12.
- Reinehr S, Becker S, Eberle A, Grether-Beck S, Häussinger D. Involvement of NADPH oxidase isoforms and SRC family kinases in CD95-dependent hepatocyte apoptosis. J Biol Chem. 2005; 280:27179–94.
- Selleri C, Sato T, Raiola AM, Rotoli B, Young NS, Maciejewski JP. Induction of nitric oxide synthase is involved in the mechanism of FAS-mediated apoptosis in hematopoietic cells. Br J Hematol. 1997;99:481–9.
- 139. Krych-Madej, J, Gebicka L. Interactions of nitrite with catalase: Enzyme activity and reaction kinetic studies. J Inorg Biochem. 2017;17:10–7.
- 140. Imai H, Nakagawa Y. Biological significance of phospholipid hydroperoxide glutathione peroxidase (PHGPx, GPx4) in mammalian cells. Free Radic Biol Med. 2003;34:145–69.
- Bauer G. Central signaling elements of intercellular reactive oxygen/nitrogen species-dependent induction of apoptosis in malignant cells. Anticancer Res. 2017;37:499–514.
- 142. Zorov DB, Juhaszova M, Sollott SJ. Mitochondrial ROS-induced ROS release: An update and review. Biochem Biophys Acta. 2006;1757:509–17.
- 143. Pletjushkina OY, Fetisova EK, Lyamzaev KG, Ivanova OY, Domnina LV, Vysskikh MY, Pustovidko AV, Vasiliev JM, Murphy MP, Chernyak BV, Skulachev VP. Long-distance apoptotic killing of cells is mediated by hydrogen peroxide in a mitochondrial ROS-dependent fashion. Cell Death Differen. 2005;12:1442–4.
- 144. Chiang CLL, Ledermann JA, Rad AN, Katz DR, Chain BM. Hypochlorous acid enhances immunogenicity and uptake of allogeneic ovarian tumor cells by dendritic cells to cross-prime tumor-specific T cells. Cancer Immunol Immunother. 2006;55:1384–95.
- 145. Chiang CLL, Ledermann JA, Aitkens E, Benjamin E, Katz DR, Chain BM. Oxidation of ovarian epithelial cancer cells by hypochlorous acid enhances immunogenicity and stimulates T cells that recognize autologous primary tumor. Clin Cancer Res. 2008;14:4898–907.
- 146. Chiang CLL, Coukos G, Kandalaft LE. Whole tumor antigen vaccines: Where are we? Vaccines. 2015;3:344–72.
- 147. Prokopowicz ZM, Arce F, Biedron R, Chiang CLL, Ciszek M, Katz DR, Nowakowska M, Zapotoczny S, Marcinkiewicz J, Chain BM. Hypochlorous acid: A natural adjuvant that facilitates antigen processing, cross-priming, and the induction of adaptive immunity. J Immunol. 2010;184:824–35.

148. Biedron R, Konopinski K, Marcinkiewicz J, Jozefowski S. Oxidation by neutrophil-derived HOCl increases immunogenicity of proteins by converting them into ligands of several endocytic receptors involved in antigen uptake by dendritic cells and macrophages. PLOS ONE. 2015;10:e01123293.

- 149. Zhou R, Huang W-L, Ma C, Zhou Y, Yao Y-Q, Wang Y-X, Gou L-T, Chen Y, Yang J-L. HOCl oxidation-modified CT26 cell vaccine inhibits colon tumor growth in a mouse model. Asian Pacific J Cancer Preven. 2012;13:4037–43.
- Held AM, Halko DJ, Hurst JK. Mechanisms of chlorine oxidation of hydrogen peroxide. J Am Chem Soc. 1978;100:5732–40.
- 151. Miyamoto S, Martinez GR, Rettori D, Augusto O, Medeiros MHG, Di Mascio P. Linoleic acid hydroperoxide reacts with hypochlorous acid, generating peroxyl radical intermediates and singlet molecular oxygen. Proc Natl Acad Sci USA. 2006;103:293–8.
- 152. Miyamoto S, Ronsein GE, Prado FM, Uemi M, Correa TC, Toma IN, Bertolucci A, Oliviera MCB, Motta FD, Medeiros MHG, Di Mascio P. Biological hydroperoxides and singlet molecular oxygen generation. IUMB Life. 2007;59:322–31.
- Sies H. Role of metabolic H₂OGeneration, redox signaling and oxidative stress. J Biol Chem. 2014; 289:8735–41.
- 154. Sies H. Hydrogen peroxide as a central redox signaling molecule in physiological oxidative stress: Oxidative eustress. Redox Biol. 2017;11:613–9.