Interactions between a Plasma-Activated Medium and Cancer Cells

Hiromasa Tanaka,* Masaaki Mizuno, Fumitaka Kikkawa, & Masaru Hori

Nagoya University, Nagoya, Japan

*Address all correspondence to: Hiromasa Tanaka, Furo-cho, Chikusa-ku, Nagoya 464-8603, Japan, E-mail: htanaka@plasma.engg.nagoya-u.ac.jp

ABSTRACT: A plasma-activated medium (PAM) is a medium that is irradiated with plasma; PAMs have garnered much attention from the plasma medicine community. PAMs induce apoptosis in many types of cancer cells and have a clinical benefit, especially in disseminated cancers such as peritoneal disseminated ovarian cancer and gastric cancer. Although the effectiveness of PAMs has been extensively reported, the mechanisms of the selective killing of cancer cells by PAMs still remain to be elucidated. Systems biology is a key to understanding the origins of the differences in cellular sensitivities to PAM.

I. INTRODUCTION

The application of nonthermal plasma to medicine was a big trend at the beginning of the 21st century; this field of medicine is often called plasma medicine.¹⁻⁶ Many *in vitro* and *in vivo* studies have demonstrated that such plasma is applicable to wound healing, blood coagulation, and cancer treatments.⁷⁻¹³ It is important to understand the mechanisms of plasma treatment; however, the complexity of both plasma and biological systems makes it very difficult to understand these mechanisms. Some research groups in the plasma medicine community have started to consider applying the concept of systems biology to plasma medicine in order to better understand the mechanisms through which nonthermal plasma acts. In this article we introduce our perspective regarding how systems biology is useful for understanding plasma medicine.

A plasma-activated medium (PAM) is a medium that is irradiated with plasma. Components of PAMs are complex, and the mechanisms of physiological outputs by PAMs are not fully understood. It is believed that reactive species generated by plasma are key players that interact with cells and tissues. Understanding the mechanisms at work in this complex system will provide insights into improving effectiveness and safety when applying PAMs for various diseases.

II. PAM AS A TOOL FOR CANCER CHEMOTHERAPY

PAMs have broadened the ways in which nonthermal plasma can be applied, by providing a tool for cancer chemotherapy. ^{14,15} In traditional cancer chemotherapy antitumor drugs attack fast-dividing cancer cells; however, they also attack fast-dividing healthy cells. By contrast, a PAM selectively killed glioblastoma brain tumor cells rather than normal astrocytes. ¹⁶ The survival and proliferation signaling network is constitutively activated in glioblastoma, which leads to the inhibition of apoptosis and promotes cell

Tanaka et al.

growth. A PAM downregulated the survival and proliferation signaling network to induce apoptosis in glioblastoma brain tumor cells. ¹⁷ *In vivo* and *ex vivo* studies have also demonstrated that both plasma and the PAM generated by our previously reported plasma source are prominent tools for cancer therapy. ^{18–20} The effectiveness and safety of PAMs will be addressed in the future.

A PAM is considered a complex cocktail of products that are generated by interactions between plasma and air, plasma and water, and plasma and molecules in air and water. Plasma interacts with oxygen, nitrogen, and water in humid air to produce molecules such as oxygen radicals, nitrogen radicals, nitric oxide (NO), and hydroxyl radicals, and it moves into the liquid phase to generate molecules such as hydrogen peroxide (H₂O₂), nitrites, and nitrates.²¹ NO is a well-known second messenger (a key player in signal transduction inside a cell), and H₂O₂ has also been recently recognized as a second messenger.^{22,23} The combined data suggest that the mechanisms of selective killing of cancer cells by a PAM might be partly explained by the biology of free radicals.²⁴ However, recent studies in plasma medicine indicated that inputs from plasma or PAMs are complicated, and it will be necessary to combine diagnostics of plasma, studies of plasma and liquids, computational simulations, and molecular biology to completely understand the mechanisms of physiological outputs by plasma and PAMs.

III. THE USE OF SYSTEMS BIOLOGY FOR UNDERSTANDING INTERACTIONS BETWEEN PLASMA AND CANCER CELLS

There are several key players in the response of a cell to various extracellular stimuli (Fig. 1). On the plasma membrane, which is made of phospholipid bilayers, there are many types of receptors, such as ion channels, cell adhesion molecules, and glycoproteins. For example, the epidermal growth factor receptor binds extracellular epidermal growth factor to transduce signals into a cell.²⁵ Ion channels function as controllers that balance the concentrations of intracellular and extracellular ions.²⁶ Epithelial cells are connected by cell adhesion molecules.²⁷ Glycoproteins play important roles in cell recognition.²⁸ There may also be some membrane proteins whose functions are unknown. Underneath the plasma membrane are cortical actin cytoskeletons, which, together with actin-binding proteins, control cellular morphology and cellular dynamics.

Inside the plasma membrane there are several second messengers, such as cyclic adenosine monophosphate, cyclic guanosine monophosphate, calcium ions, phosphatidylinositols, and NO. NO is an important cellular signaling molecule that is involved in various physiological processes, including vasodilatation, neurotransmission, and protection against infection.²⁹ NO is synthesized from arginine by NO synthases in a cell, has a lifetime of only a few seconds, and freely penetrates cell membranes. NO is synthesized in the endothelium of blood vessels and diffuses into smooth muscles to react with guanylate cyclase and increase cyclic guanosine monophosphate, which results in the relaxation of smooth muscle and an increase in blood flow. NO has been reported to induce apoptosis in some cells and inhibit apoptosis in other cells.³⁰ Thus, high

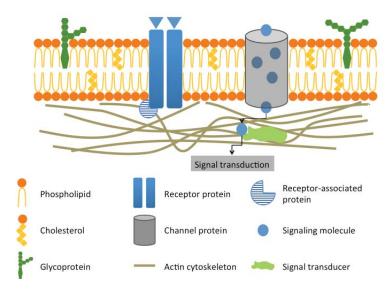


FIG. 1: Key players in cellular responses to extracellular stimuli. Various types of proteins on the plasma membrane receive and sense different extracellular environments. Actin cytoskeletons underlie the plasma membrane to form and change cell morphology. Inside the cell, signaling molecules such as nitric oxide and hydrogen peroxide are generated by extracellular signals for signal transduction.

concentrations of NO (more than ~1 µmol/L), or prolonged exposure to NO, induces apoptosis in some cells through various molecular mechanisms, whereas physiological concentrations of NO (less than ~100 nmol/L) protect cells from apoptosis. H₂O₂ is a well-known reactive oxygen species and has recently been recognized as a second messenger. Various growth factors and cytokines generate H_2O_2 in a cell through activation of nicotinamide adenine dinucleotide phosphate hydrogen oxidases. Low concentrations of H_2O_2 (less than ~10 µmol/L) induce antioxidants that protect a cell from oxidative stress, whereas high concentrations of H_2O_2 (more than ~10 µmol/L) induce apoptosis in a cell. In summary, both NO and H_2O_2 exert concentration-dependent and cell-type-specific effects; however, the mechanisms by which a cell senses and regulates the concentrations of these species are unclear.

The survival and proliferation signaling network is also modulated by NO and $\rm H_2O_2$. For example, a key molecule in this signaling network, the phosphatase and tensin homolog (PTEN), is reversibly inactivated by $\rm H_2O_2$ through oxidation of the essential cysteine residue of its active site.³³ PTEN is also inactivated by NO through S-nitrosylation, which is a covalent modification of cysteine residues that induces degradation of the PTEN protein via ubiquitination.³⁴

Systems biology is a field of study in which biological phenomena are studied as a system, and it combines classical molecular biology, biochemistry, computational science, and systems engineering.³⁵ Biological systems receive various inputs from their environments and show several physiological outputs. The relationships between inputs

Tanaka et al.

and outputs are determined by the structure of the signaling pathways. For example, the survival and proliferation signaling network has several negative feedback loops and amplifiers (Fig. 2), and mutations in this signaling network can cause changes in the strength of such regulatory functions.³⁶ In this way, differences in the sensitivities of cells with different mutations to various stimuli can be explained by differences in the structure of the signaling pathways.

Cellular sensitivities to PAMs are determined by the structure of the signaling pathways that regulate the survival and proliferation signaling network. When levels of NO and H_2O_2 that are higher than physiological levels are generated in a cell, the cell switches on apoptotic signals. The doses of NO and H_2O_2 that induce apoptosis depend on the cell type. Thus the different sensitivity of different cell types to a PAM might be explained by the structure of the signaling pathways of the survival and proliferation signaling network. Such system analyses are needed to understand the mechanisms of the selective killing of cancer cells by PAMs.

IV. CONCLUSION

We propose PAMs as a new type of cancer chemotherapy. A PAM potentially generates H_2O_2 and NO in a cell. Both NO and H_2O_2 are key signaling molecules in several sig-

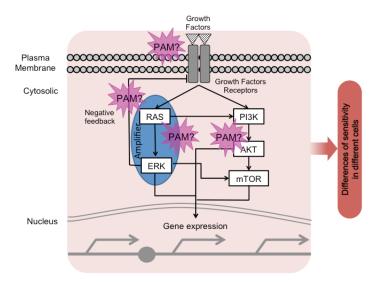


FIG. 2: Structure of signaling pathways in the survival and proliferation signaling network. There are several negative feedback loops and amplifiers in the signaling pathways of the survival and proliferation signaling networks. Differences in the sensitivities of different cell types to a plasma-activated medium (PAM) might be explained by changes in the strength of the structures of these signaling pathways. mTOR, mammalian target of rapamycin; PI3K, phosphoinositide 3-kinase.

naling networks and also interact with the survival and proliferation signaling network through S-nitrosylation and the oxidation of cysteine residues of the PTEN protein. Differences in the sensitivities of various cells to a PAM might be explained by differences in the structure of the signaling pathways in the survival and proliferation signaling network. In the future, interactions between plasma/PAMs and cells/tissues will be understood better using systems biology.

ACKNOWLEDGMENTS

This work was partly supported by a Grant-in-Aid for Scientific Research on Innovative Areas ("Plasma Medical Innovation" grant nos. 24108002 and 24108008), a Grant-in-Aid for Young Scientists (A) (grant no. 15H05430), and a Grant-in-Aid for Challenging Exploratory Research (grant no. 15K13390) from the Ministry of Education, Culture, Sports, Science and Technology of Japan.

REFERENCES

- 1. Fridman G, Friedman G, Gutsol A, Shekhter AB, Vasilets VN, Fridman A. Applied plasma medicine. Plasma Process Polym. 2008;5:503–33.
- 2. Laroussi M. Low-temperature plasmas for medicine? IEEE Trans Plasma Sci. 2009;37:714–25.
- 3. Weltmann KD, Kindel E, von Woedtke T, Hahnel M, Stieber M, Brandenburg R. Atmospheric-pressure plasma sources: prospective tools for plasma medicine. Pure Appl Chem. 2010;82:1223–37.
- 4. Kong MG, Kroesen G, Morfill G, Nosenko T, Shimizu T, van Dijk J, Zimmermann JL. Plasma medicine: an introductory review. New J Phys. 2009;11:115012.
- 5. Morfill GE, Kong MG, Zimmermann JL. Focus on plasma medicine. New J Phys. 2009;11:115011.
- von Woedtke T, Reuter S, Masur K, Weltmann KD. Plasmas for medicine. Phys Rep. 2013;530: 291–320.
- Keidar M, Walk R, Shashurin A, Srinivasan P, Sandler A, Dasgupta S, Ravi R, Guerrero-Preston R, Trink B. Cold plasma selectivity and the possibility of a paradigm shift in cancer therapy. Br J Cancer. 2011;105:1295–301.
- 8. Vandamme M, Robert E, Pesnel S, Barbosa E, Dozias S, Sobilo J, Lerondel S, Le Pape A, Pouvesle JM. Antitumor effect of plasma treatment on U87 glioma xenografts: preliminary results. Plasma Process Polym. 2010;7:264–73.
- 9. Volotskova O, Hawley TS, Stepp MA, Keidar M. Targeting the cancer cell cycle by cold atmospheric plasma. Sci Rep. 2012;2:636.
- Lee HJ, Shon CH, Kim YS, Kim S, Kim GC, Kong MG. Degradation of adhesion molecules of G361 melanoma cells by a non-thermal atmospheric pressure microplasma. New J Phys. 2009;11:115026.
- 11. Gherardi M, Turrini E, Laurita R, Gianni ED, Ferruzzi L, Liguori A, Stancampiano A, Colombo V, Fimognari C. Atmospheric non-equilibrium plasma promotes cell death and cell-cycle arrest in a lymphoma cell line. Plasma Process Polym. 2015;12:1354–63.
- 12. Gweon B, Kim D, Kim DB, Jung H, Choe W, Shin JH. Plasma effects on subcellular structures. Appl Phys Lett 2010;96:101501.
- 13. Ishaq M, Kumar S, Varinli H, Han ZJ, Rider AE, Evans MD, Murphy AB, Ostrikov K. Atmospheric gas plasma-induced ROS production activates TNF-ASK1 pathway for the induction of melanoma cancer cell apoptosis. Mol Biol Cell. 2014;25:1523–31.
- Tanaka H, Mizuno M, Ishikawa K, Takeda K, Nakamura K, Utsumi F, Kajiyama H, Kano H, Okazaki Y, Toyokuni S, Maruyama S, Kikkawa F, Hori M. Plasma medical science for cancer therapy: toward cancer therapy using nonthermal atmospheric pressure plasma. IEEE Trans Plasma Sci. 2014;42:3760–4.

Tanaka et al.

 Tanaka H, Mizuno M, Toyokuni S, Maruyama S, Kodera Y, Terasaki H, Adachi T, Kato K, Kikkawa F, Hori M. Cancer therapy using non-thermal atmospheric pressure plasma with ultra-high electron density. Phys Plasmas. 2015;22:122003.

- Tanaka H, Mizuno M, Ishikawa K, Nakamura K, Kajiyama H, Kano H, Kikkawa F, Hori M. Plasmaactivated medium selectively kills glioblastoma brain tumor cells by down-regulating a survival signaling molecule, AKT kinase. Plasma Med. 2011;1:265–77.
- Tanaka H, Mizuno M, Ishikawa K, Nakamura K, Utsumi F, Kajiyama H, Kano H, Maruyama S, Kikkawa F, Hori M. Cell survival and proliferation signaling pathways are downregulated by plasmaactivated medium in glioblastoma brain-tumor cells. Plasma Med. 2012;2:207–20.
- Utsumi F, Kajiyama H, Nakamura K, Tanaka H, Mizuno M, Ishikawa K, Kondo H, Kano H, Hori M, Kikkawa F. Effect of indirect nonequilibrium atmospheric pressure plasma on anti-proliferative activity against chronic chemo-resistant ovarian cancer cells in vitro and in vivo. Plos One. 2013;8:e81576.
- Okazaki Y, Wang Y, Tanaka H, Mizuno M, Nakamura K, Kajiyama H, Kano H, Uchida K, Kikkawa F, Hori M, Toyokuni S. Direct exposure of non-equilibrium atmospheric pressure plasma confers simultaneous oxidative and ultraviolet modifications in biomolecules. J Clin Biochem Nutr. 2014;55:207–15.
- Adachi T, Tanaka H, Nonomura S, Hara H, Kondo SI, Hori M. Plasma-activated medium induces A549 cell injury via a spiral apoptotic cascade involving the mitochondrial-nuclear network. Free rRadic Biol Med. 2014; 79C:28–44.
- Bruggeman P, Leys C. Non-thermal plasmas in and in contact with liquids. J Phys D Appl Phys. 2009;42:053001.
- 22. Reth M, Hydrogen peroxide as second messenger in lymphocyte activation. Nat Immunol. 2002;3: 1129–34.
- 23. Breton-Romero R, Lamas S. Hydrogen peroxide signaling in vascular endothelial cells. Redox Biol. 2014;2:529–34.
- 24. Toyokuni S, Okamoto K, Yodoi J, Hiai H. Persistent oxidative stress in cancer. FEBS Lett. 1995;358:1–3.
- 25. Zandi R, Larsen AB, Andersen P, Stockhausen MT, Poulsen HS. Mechanisms for oncogenic activation of the epidermal growth factor receptor. Cell Signal 2007;19:2013–23.
- 26. Bates E. Ion channels in development and cancer. Annu Rev Cell Dev Biol. 2015;31:231–47.
- Lecuit, Yap AS. E-cadherin junctions as active mechanical integrators in tissue dynamics. Nat Cell Biol. 2015;17:533–9.
- 28. Feizi T, Haltiwanger RS. Editorial overview: Carbohydrate-protein interactions and glycosylation: glycan synthesis and recognition: finding the perfect partner in a sugar-coated life. Curr Opin Struct Biol. 2015;34:vii–ix.
- 29. Zhao Y, Vanhoutte PM, Leung SW. Vascular nitric oxide: beyond eNOS. J Pharmacol Sci. 2015;129: 83–94.
- Boyd CS, Cadenas E. Nitric oxide and cell signaling pathways in mitochondrial-dependent apoptosis. Biol Chem. 2002;383:411–23.
- 31. Thomas DD, Ridnour LA, Isenberg JS, Flores-Santana W, Switzer CH, Donzelli S, Hussain P, Vecoli C, Paolocci N, Ambs S, Colton CA, Harris CC, Roberts DD, Wink DA. The chemical biology of nitric oxide: implications in cellular signaling. Free Radic Biol Med. 2008;45:18–31.
- 32. Veal EA, Day AM, Morgan BA. Hydrogen peroxide sensing and signaling. Mol Cell. 2007;26:1-14.
- 33. Lee SR, Yang KS, Kwon J, Lee C, Jeong W, Rhee SG. Reversible inactivation of the tumor suppressor PTEN by H2O2. J Biol Chem. 2002;277:20336–42.
- 34. Kwak YD, Ma T, Diao S, Zhang X, Chen Y, Hsu J, Lipton SA, Masliah E, Xu H, Liao FF. NO signaling and S-nitrosylation regulate PTEN inhibition in neurodegeneration. Mol Neurodegener. 2010;5:49.
- 35. Kitano H. Systems biology: a brief overview. Science. 2002;295:1662-4.
- 36. Caron E, Ghosh S, Matsuoka Y, Ashton-Beaucage D, Therrien M, Lemieux S, Perreault C, Roux PP, Kitano H. A comprehensive map of the mTOR signaling network. Mol Syst Biol. 2010;6:453.